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| (54) Title: METHODS FOR TARGETING, ENRICHING, DETECTING AND/OR ISOLATING TARGET NUCLEIC ACID SEQUENCE USING RECA-LIKE RECOMBINASE (57) Abstract <p>A method for targeting, enriching, detecting and isolating a double-stranded nucleic acid target sequence by adding heterologous probes to the reaction system along with the reduced amount of homologous probes in the process of homologous pairing and/or strand exchange between the homologous probe and the target sequence in the presence of a RecA-like recombinase.</p> | | |

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DESCRIPTION

**METHODS FOR TARGETING, ENRICHING, DETECTING AND/OR
ISOLATING TARGET NUCLEIC ACID SEQUENCE USING RECA-LIKE
RECOMBINASE**

5 **Background of the Invention****Area of the Art**

The invention relates generally to methods for targeting, enriching, detecting and/or isolating target nucleic acid sequence using RecA-like recombinase, and specifically to methods for targeting, enriching, detecting and/or isolating double-
10 stranded nucleic acid target sequence using RecA-like recombinase in the presence of both a homologous probe and a heterologous probe.

Description of the Prior Art

A variety of recombinases, which catalyze in vitro homologous pairing and/or exchange of DNA strands, have been isolated from various prokaryotes and
15 eukaryotes. Among these recombinases, RecA protein, a recombinase derived from *Escherichia coli*, has been extensively investigated. (Shibata T., Cell Technology, 9, No. 4, 281-291 (1990)). RecA protein is known to catalyze in vitro homologous pairing of single-stranded DNA with double-stranded DNA and thus to generate homologously paired triple-stranded DNA or other triple-stranded joint DNA
20 molecules. (Rigas B., et al., Proc. Natl. Acad. Sci. USA, 83: 9591-9595 (1986); Hsieh P. et al., Proc. Natl. Acad. Sci. USA, 89: 6492-6496 (1992); Ferrin L. J. et al., Science, 254: 1494-1497 (1991), etc.). RecA protein is also reported to catalyze the formation of a four-stranded DNA structure known as a double D-loop. In this reaction, two types of complimentary single-stranded DNA are used as homologous
25 probes to target double-stranded DNA, which has a homologous site for the single-stranded DNA probe. (Sena E. P., Nature Genetics, 3: 365-372 (1993); Jayasena V. K. et al., J. Mol. Biol., 230: 1015 (1993)). In addition to DNA-DNA hybridization, RecA protein can also promote RNA-DNA hybridization. For example, single-stranded DNA coated with RecA protein can recognize complementarity with naked RNA
30 (Kirkpatrick and Radding, 1992; Kirkpatrick et al., 1992).

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By utilizing the property of RecA protein, methods have been developed for isolating specific double-stranded target DNA existing in a solution at a very low level (at a molar ratio of 1:50 molecules to 1 : several hundred molecules) (Rigas B. et al., Proc. Natl. Acad. Sci. USA, 83: 9591-9595 (1986); Teintze M. et al., Biochem. Biophys. Res. Commun., 211: 804-811 (1995); US Patent No. 4,888,274). In situ hybridization methods have also been developed for detecting double-stranded target DNA in a fixed cell (WO 93/05177). These methods use RecA to mediate homologous pairing between the target DNA and a homologous probe containing a sequence sufficiently complementary to the target DNA to form a homologous probe/target DNA complex.

Homologous pairing that is catalyzed by a recombinase, such as RecA protein, leads to the formation of networks (coaggregates) comprising RecA proteins, total DNA (target DNA + heterologous DNA) and homologous probes in the system (Tsang S. S. et al., Biochemistry, 24: 3226-3232 (1985); Gonda D. K. et al., J. Biol. Chem., 261: 13087-13096; Chow S. A. et al., Proc Natl. Acad. Sci. USA, 82: 5646-5650 (1985)). The efficiency of the homologous pairing occurring between double-stranded target DNA and single-stranded nucleic acid (homologous probe), which is complementary to the double-stranded target DNA and to which RecA protein is bound, is greatly reduced when an excessive amount of heterologous DNA is present in a given sample. To prevent this reduction in reaction efficiency, the amount of RecA protein, as well as that of RecA protein-coated homologous probe has to be increased in proportion to the amount of the total DNA in the sample (Rigas B. et al., Proc. Natl. Acad. Sci. USA, 83: 9591-9595 (1986)).

However, an increase in the amount of RecA protein-coated homologous probe in proportion to the total amount of DNA in a given sample increases the amount of the RecA protein/homologous probe/target DNA/heterologous DNA complex, which correspondingly increases the final amount of heterologous DNA contaminating the double-stranded target DNA recovered from the sample. Such contamination reduces the specificity of the reaction. Therefore, it seems a problem that in isolating double-stranded target DNA, the amount of the contaminating heterologous DNA recovered together with double-stranded target DNA is dependent

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on the amount of the homologous probe used in the reaction. This problem is especially significant if the ratio of target DNA to heterologous DNA is less than 1:1,000.

The heterologous DNA may be removed from the complex by utilizing the differences in sodium chloride or Mg^{2+} sensitivity between the probe/target DNA and the probe/heterologous DNA complexes (Honigberg S. M. et al., Proc. Natl. Acad. Sci. USA, 83: 9586-9590 (1986); Rigas B. et al., Proc. Natl. Acad. Sci. USA, 83: 9591-9595 (1986); US Patent No. 4,888,274). It was reported that when one uses a long, circular homologous probe of more than 5,000 nucleotides or a long stretch linear homologous probe of more than 3,000 nucleotides, it seems possible to isolate the double-stranded target DNA contained in a given sample at a ratio of 1 target DNA molecule per about 200 to 1,000 heterologous DNA molecules with certain specificity. The heterologous DNA may be removed from the complex of homologous probe/target DNA by utilizing the difference between the two complexes in sensitivity to sodium chloride or Mg^{2+} . The stability of the homologous probe/heterologous DNA complex significantly differs from that of the homologous probe/target DNA complex.

However, when one uses a circular homologous probe containing a comparatively short sequence, e.g., a 700-nucleotide sequence complementary to a portion of the target DNA, the final yield of the double-stranded target DNA is significantly lowered. This is the case even if the concentration of the double-stranded target DNA is relatively high, such as one molecule per 50 molecules in a given sample. (Teintze M. et al., Biochem. Biophys. Res. Comm., 211: 804-811 (1995)). Therefore, when the amount of the double-stranded target DNA present in the DNA sample is extremely small (for example, at a molar ratio less than 1 molecule/ 1,000 molecules), it is unclear whether or not one can use a homologous probe to isolate double-stranded target DNA.

It is therefore extremely difficult to selectively eliminate the heterologous DNA from the RecA protein/ homologous probe/ target DNA/ heterologous DNA complex, wherein the amount of heterologous DNA exceeding that of target DNA is more than 1,000-fold in a given sample. Particularly, when the length of the complementary sequence common to both the double-stranded target DNA and the homologous probe

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(the length of homologous probe) is short, i.e., less than 700 nucleotide sequence, the use of such short probes will make it more difficult for the selective elimination of heterologous DNA. This is because the bond within the homologous probe/target DNA complex, mediated by a short RecA-bound homologous probe, is significantly
5 unstable in comparison with the bond within the same complex, mediated by a long homologous probe consisting of more than 3,000 complementary nucleotide sequence to the whole region of the double-stranded target DNA. Washing under stringent conditions will break the homologous probe/target DNA complex when a short probe is used.

10 When ATP γ S is employed as a co-factor, it is difficult to selectively eliminate only the heterologous DNA from the complex, since the bond within the RecA protein/homologous probe/ heterologous DNA/ target DNA complex is extremely stable. Alternatively, it is possible to eliminate RecA proteins from the complex prior to the process of removing the heterologous DNA from it. However, the stability of the
15 homologous probe/target DNA complex without RecA protein distinctively decreases in comparison with that of the RecA protein/homologous probe/target DNA complex. This alternative, therefore, is not preferred. (Teintze M. et al., Biochem. Biophys. Res. Commun., 211: 804-811 (1995)).

Due to the fact that the amount of the contaminating heterologous DNA
20 recovered with double-stranded target DNA is dependent on the amount of the homologous probe used in the reaction, as previously described, reducing the amount of homologous probe in the reaction may facilitate the efficiency of the selective elimination of heterologous DNA. Reducing only the amount of homologous probe, however, will also reduce the efficiency of homologous pairing between a homologous
25 probe and a target DNA.

It has been demonstrated that the efficiency of the reaction, which depends on the length of the complementary sequence common to both the double-stranded target DNA and the homologous probe (the length of homologous probe), decreases as the length of the homologous probe decreases. (Hsieh P. et al., Proc. Natl. Acad. Sci.
30 USA, 89: 6492 (1992); Sena E. P., Nature Genetics, 3: 365-372 (1993); Jayasena V. K. et al., J. Mol. Biol., 230:1015 (1993), etc.). Particularly, the efficiency of the

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homologous pairing is remarkably reduced when a homologous probe of less than several hundred nucleotide sequence is used in the reaction. Accordingly, it is extremely difficult to detect and/or isolate double-stranded target DNA efficiently by using a short homologous probe alone, particularly, when the amount of heterologous DNA existing in a given sample is more than 1,000-fold amount of target DNA.

It was reported that when one uses a long circular homologous probe of more than 5,000 nucleotides in homologous pairing between the homologous probe and a target DNA, contained at a ratio of one target DNA molecule to 1,000 heterologous DNA molecules in a sample, the reaction efficiency will be reduced when the amount of the RecA-coated homologous probe is reduced. The reduction of the reaction efficiency of homologous pairing, however, was suppressed by adding a long circular heterologous probe of more than 6,000 nucleotides to the pairing reaction at the ratio of less than sevenfold (molar ratio) of the amount of the homologous probe (Honigberg S. M. et al., Proc. Natl. Acad. Sci. USA, 83: 9586-9590 (1986)).

Therefore, a need exists to improve the efficiency and the specificity of the reaction of the conventional methods using a recombinase such as RecA protein for targeting, enriching, detecting and/or isolating an extremely small amount of double-stranded target DNA present in a given DNA sample (for example, at a molar ratio of less than one molecule to 1,000 molecules) (M. Teintze et al, Biochem. Biophys. Res. Comm., 211, 804-811 (1995)). A need also exists for employing a short-strand homologous probe for various applications as discussed above.

Summary of the Invention

It is therefore an object of the present invention to provide a method utilizing RecA-like recombinase for targeting, enriching, detecting, and/or isolating a target nucleic acid sequence with high efficiency and specificity. It is also an objective of the present invention to provide a RecA-like recombinase mediated method utilizing a short-strand probe for targeting, enriching, detecting, and/or isolating a low level target nucleic acid sequence in a given sample.

These and other objects and advantages are achieved by using a method of the present invention utilizing RecA-like recombinase for targeting, enriching, detecting and/or isolating a double-stranded nucleic acid target sequence in a sample. The method comprises the steps of providing a RecA-like recombinase, a homologous nucleic acid probe, a heterologous nucleic acid probe; and mixing the RecA-like recombinase, the homologous nucleic acid probe, the heterologous nucleic acid probe with the double-stranded target nucleic acid sequence in the sample, wherein the specificity of the homologous pairing and/or strand exchange between the double-stranded nucleic acid target sequence and the homologous nucleic acid probe is increased due to the addition of the heterologous nucleic acid probe.

The methods in accordance with the present invention have been found to provide a number of advantages. As explained in greater detail below, it has been found that the present invention makes it possible to target, enrich, detect, and/or isolate a low-level target nucleic acid sequence contained in a given sample with high efficiency and specificity. The present invention also makes it possible to use a short homologous probe to detect a target sequence having only partial sequence been revealed.

The methods in accordance with the present invention are well suited for use in isolation and subsequent cloning of a target gene from a mixture of cDNAs or genomic DNAs. They are also well suited for use in screening of the target gene from various gene libraries, particularly, isolating and screening of the target gene whose nucleotide sequence or amino acid sequence is only partially revealed.

The methods of the present invention may also be used in the amplification of the target DNA sequence using RecA-like recombinase; mapping of various genes,

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such as RARE-based mapping using an oligonucleotide probe; nucleotide sequence specific modification or cleavage of the target DNA using an oligonucleotide.

Furthermore, the methods of the present invention may be used in in-situ hybridization methods using RecA-like recombinase to improve the specificity or the efficiency of the hybridization. They may also be used as a gene therapy technique by means of gene alternation or transcription inhibition using in vivo gene-targeting in living cells.

In this use of the method in accordance with the present invention, homologous pairing and/or strand exchange between a homologous probe and a target sequence, mediated by RecA-like recombinase, occurs in the presence of a RecA-like recombinase coated heterologous probe. In an embodiment of the present invention, the weight ratio of the amount of homologous probe to the amount of heterologous probe is between about 1:1 to 1:500.

The invention is defined in its fullest scope in the appended claims and is described below in its preferred embodiments.

Detailed Description of the Invention

The present invention is based on a surprise discovery that the addition of a heterologous probe to a RecA-like recombinase mediated reaction system increases the specificity of homologous pairing and/or strand exchange between a target DNA and a homologous probe complimentary to the target DNA, particularly, when a short homologous probe is used.

In addition, it is a surprise discovery of the present invention that reducing the amount of a homologous probe and adding a heterologous probe instead into a RecA-like recombinase mediated reaction system provide a higher specificity than that obtained when the homologous probe is used alone. This is also true even when a short linear homologous probe of less than several hundred nucleotides in length is used in the presence of at least 1,000-fold excess of heterologous DNA over the target DNA in a given sample.

Accordingly, the present invention provides a method for targeting, enriching, detecting and/or isolating double-stranded nucleic acid target sequence in a sample by using RecA-like recombinase. The method comprises the steps of providing a RecA-like recombinase, a homologous nucleic acid probe, a heterologous nucleic acid probe; and mixing the RecA-like recombinase, the homologous nucleic acid probe, the heterologous nucleic acid probe with the double-stranded target nucleic acid sequence in the sample, wherein the specificity of the homologous pairing and/or strand exchange between the double-stranded target nucleic acid sequence and the homologous nucleic acid probe is increased due to the addition of the heterologous nucleic acid probe.

For the purpose of the present invention, the "efficiency" is determined based on the amount of a target nucleic acid sequence that is targeted, enriched, detected, or isolated. The "specificity" is determined based on the ratio of the amount of the target nucleic acid sequence that is targeted, enriched, detected, or isolated and the total amount of the target nucleic acid sequence plus the heterologous nucleic acid sequence that is targeted, enriched, detected, or isolated.

RecA-like recombinases utilized in the present invention include recombinases which have catalytic activity similar to of RecA protein derived from *Escherichia coli*.

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RecA protein can mediate both homologous pairing and/or strand exchange between appropriate DNA molecules in in vitro homologous recombination assays (Kowalczykowski, S., *Ann. Rev. Biochem. Biophysical Chem.*, 20:539-575 (1991), Radding C., M., *Biochem. Biophys. Acta* 1008:131-139 (1989), Radding C., M., *J. Biol. Chem.* 266:5355-5358 (1991); also see Golub, E., et al., *Nucleic Acids Res.* 20:3121-3125 (1992)). In addition to DNA-DNA hybridization, RecA protein can promote RNA-DNA hybridization. For example, RecA protein coated single stranded DNA can recognize complementarity with naked RNA (Kirkpatrick, S. et al., *Nucleic Acids Res.* 20:4339-4346 (1992)). Therefore, any recombinase which can promote both homologous pairing and/or strand exchange between appropriate DNA molecules or between DNA and RNA molecules may be used in the present invention.

RecA-like recombinases have been isolated and purified from many prokaryotes and eukaryotes. The examples of such recombinases include, but are not limited to, the wild type RecA protein derived from *Escherichia coli* (Shibata T. et al., *Method in Enzymology*, 100:197 (1983)), and mutant types of the RecA protein (e.g., RecA 803: Madiraju M. et al., *Proc. Natl. Acad. Sci. USA*, 85: 6592 (1988); RecA 441(Kawashima H. et al., *Mol. Gen. Genet.*, 193: 288 (1984), etc.); uvsX protein, a T4 phage-derived analogue of the protein (Yoncsaki T. et al., *Eur. J. Biochem.*, 148: 127 (1985)); RecA protein derived from *Bacillus subtilis* (Lovett C. M. et al., *J. Biol. Chem.*, 260: 3305 (1985)); RecI protein derived from *Ustilago* (Kmiec E. B. et al., *Cell*, 29 :367 (1982)); RecA-like protein derived from heat-resistant bacteria (such as *Thermus aquaticus* or *Thermus thermophilus*) (Angov E. et al., *J. Bacteriol.*, 176: 1405 (1994); Kato R. et al., *J. Biochem.*, 114: 926 (1993)); and RecA-like protein derived from yeast, mouse and human (Shinohara A. et al., *Nature Genetics*, 4: 239 (1993)).

In a preferred embodiment, RecA protein isolated and purified from a culture of *Escherichia coli* may be used. (E.g., Kuramitsu S. et al., *J. Biochem.*, 90: 1033 (1981); Shibata T. et al., *Methods in Enzymology*, 100: 197 (1983)). Commercially available RecA protein may also be used (Boehringer-Mannheim, Pharmacia).

As used herein, the term "nucleic acid" encompasses RNA as well as DNA and cDNA. In the present invention, there is no specific restriction on a double-stranded

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nucleic acid target sequence for its length, type, or shape. For example, the double-stranded nucleic acid target sequence may be in a circular form or a linear form.

Preferably, the double-stranded nucleic acid target sequence is double-stranded target DNA. The double-stranded target DNA may be, but is not limited to, genomic
5 DNA, cDNA derived from prokaryotes and eukaryotes, DNA derived from viruses or bacteriophages, fragments of such genomic DNA or cDNA, and various species of such DNA contained in various kinds of DNA libraries.

A double-stranded nucleic acid target sequence of the present invention may be a nucleic acid sequence contained in a solution; in fixed cells, cellular structures, and
10 intercellular structures; or in living cells or cellular structures which are not fixed. Examples of cells, cellular structures and intercellular structures include, but are not limited to, bacteria, viruses, cellular organs such as nucleus, mitochondria, or chromosome, and parasites such as viruses or bacteria existing in biological samples such as blood. Cells or cellular structures may be fixed by means of conventional
15 methods using organic solvents (e.g., methanol, ethanol etc.), acids (e.g., acetic acid) or cross-linking agents (e.g., formalin, paraformaldehyde). Those methods of fixing are well within the skill of the art in view of the instant disclosure.

A double-stranded nucleic acid target sequence of the present invention, if desired, may be labeled using conventional methods well known in the art. The target
20 sequence may be labeled by, but not limited to, radio isotopes (e.g., ^{32}P , ^{35}S , etc.), fluorescent pigments (e.g., FITC, rhodamine etc.), enzyme labels (e.g., peroxidase, alkaline phosphatase etc.), chemiluminescent agents (e.g., acridinium ester etc.), and a various sorts of labels and ligands such as biotin or digoxigenin. Those labeling methods are well known to a person skilled in the art in view of the instant disclosure.

25 It should be understood that, although the methods in accordance with the present invention are primarily directed to the targeting, enriching, detecting, and/or isolating of double-stranded target DNA, they may also be used for other duplex nucleic acid target sequences. Examples of such duplex nucleic acid target sequences include, but are not limited to, DNA/RNA sequences or possibly double-stranded RNA
30 elements, double-stranded or single-stranded nucleic acid sequences associated with viral, bacterial or parasitic pathogens.

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As used herein, the term "single-stranded nucleic acid probe" means single-stranded nucleic acids. The term "nucleic acid" encompasses RNA as well as DNA and cDNA. Preferably, single-stranded DNA is employed. There is no restriction regarding the shape of a probe, and thus either a circular form or a linear form may be used.

Commercially available products of single-stranded or double-stranded nucleic acid sequences can be used as probes of the present invention. Probes can also be prepared using the methods well-known in the art. For example, probes may be prepared directly from viruses, bacteriophages, plasmids, cosmids, or other vectors which have a target sequence, by means of, e.g., a nick-translation method and random prime method or the like. If desired, a probe can be prepared by restriction digest of the segment corresponding to the probe from the vector and followed by electrophoretic isolation of the specific restriction fragment, or by amplification of the probe sequence using the PCR method. Although the probe thus obtained is usually double-stranded, if desired, a single-stranded probe can be obtained by denaturing or by sub-cloning of the double-stranded sequence into a single-stranded vector such as M13 phage. Alternatively, a single-stranded probe can be prepared by oligo-nucleotide synthetic methods. A long probe can be prepared by joining sub-fragments of the probe after the synthesis of those sub-fragments.

A homologous probe in accordance with the present invention is a nucleic acid probe which has a region of homology with a selected base sequence in a duplex nucleic acid target sequence. Preferably, a homologous probe is a single-stranded DNA probe. As used herein, the term probe "homology" with the target means that the single-stranded probe and target duplex have a region of similar or exact base pair sequence which allows the probe to recognize and hybridize with the corresponding base pair region in the duplex target. The extent of base pair mismatching which is allowed without losing homology may be as high as 20% to 30%, depending on the distribution and lengths of mismatched base pairs. In a preferred embodiment, the homologous probe is single-stranded nucleic acid which contains the sequence having at least more than 70% homology to the partial or whole sequence of a target nucleic acid. In order to ensure sequence specific homologous pairing (hybridization reaction)

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between the double-stranded nucleic acid target sequence and the homologous probe, it is preferable that the homologous probe generally contains the sequence that is at least 90-95% homologous to the partial or whole sequence of the double-stranded nucleic acid target sequence.

5 A homologous probe in accordance with the present invention may be prepared by denaturing a double-stranded nucleic acid probe which is complementary to either one or both strands of a target sequence. The homologous probe may also contain an extended terminal portion which is not complementary to any of the nucleic acid strands in the sample. When both strands of a double-stranded homologous probe
10 have such extended sequences at their termini, the extended sequences may be complementary to each other.

 A homologous probe in accordance with the present invention is at least 15 nucleic acid residues in length, and preferably 15-2,000 nucleic acid residues in length. A longer polynucleotide probe (more than 2,000 nucleic acid in length) may also be
15 used (Hsieh P. et al., Proc. Natl. Acad. Sci. USA, 89:6492 (1992)). Preferably, the length of required homology between a probe and a target is at least about 15 base pairs.

 If desired, a homologous probe may be labeled for detection and/or isolation using conventional methods known in the art. Examples of the labels that may be used
20 to label a homologous probe include, but are not limited to, radio isotopes (e.g., ³²P, ³⁵S etc.), fluorescent pigments (e.g., FITC, rhodamine etc.), enzyme labels (e.g., peroxidase, alkaline phosphatase etc.), chemiluminescent agents (e.g., acridinium ester etc.), and a various sorts of labels and ligands such as biotin or digoxigenin.

 In the present invention, at least one species of the homologous probes is used
25 in combination with at least one species of RecA-like recombinase. For example, a homologous probe may be coated with RecA-like recombinase to form a homologous probe/RecA-like recombinase complex using a method that is well known in the art. The amount of homologous probe to be used in a sample is usually 0.1-200ng, preferably 0.5-150ng, when the sample contains about 1 µg of DNA (target DNA and
30 heterologous DNA).

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The heterologous probe employed in the present invention is a nucleic acid probe which is not sufficiently complementary to the target sequence. A heterologous probe is not sufficiently complementary to a target sequence if it cannot hybridize with the target sequence nor form a stable heterologous probe/target sequence complex. In a preferred embodiment, the sequence of the heterologous probe may show low complementarity to a sequence other than the target sequence in a target molecule comprising the target sequence and a vector sequence. For example, it may show low complementarity to a vector sequence, in which the target sequence is integrated or is used for constructing the gene library. However, it is preferred that such complementarity should be sufficiently low such that a heterologous probe/target molecule complex or a heterologous probe/vector sequence complex would not be formed.

The heterologous probe may be a DNA probe as well as an RNA probe. Preferably, the heterologous probe is single-stranded DNA. There is no restriction regarding the shape of a probe, and thus either a circular form or a linear form can be used. The probe may be prepared by denaturing the double-stranded heterologous probe.

For example, if the DNA sample is derived from a human, the heterologous probe used in the system may be a single-stranded nucleic acid probe derived from, but not limited to, a virus or bacteriophage, preferably, single-stranded phage DNA such as M13 or ϕ X174, or single-stranded DNA generated from the fragment of lambda phage DNA; a single-stranded nucleic acid probe derived from microorganisms, such as RNA derived from *Escherichia coli* or yeast; a single-stranded nucleic acid probe derived from eukaryotes except humans, such as single-stranded DNA generated from DNA fragments from salmon sperm.

Preferably, the heterologous probe employed in the present invention does not contain any labels or ligands.

The heterologous probe used in the present invention is at least 15 nucleic acid residues in length. Preferably it is about 15-20,000 nucleic acid residues in length, and more preferably 60-10,000 nucleic acid in length. A longer polynucleotide probe, such as a probe of more than 20,000 nucleic acid residues in length, may also be used.

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In the present invention, at least one species of heterologous probes is used in combination with at least one species of recombinase. For example, a heterologous probe of the present invention may be coated with RecA-like recombinase to form a heterologous probe/RecA-like recombinase complex under reaction conditions well known in the art. The amount of a heterologous probe to be used in a sample is usually 0.1-500 ng, preferably 20-250 ng, when the sample contains about 1 μ g of DNA (target DNA and heterologous DNA).

In the present invention, co-factors may be added to the reaction system in association with the recombinase used. For example, when RecA protein derived from *Escherichia coli* is used as a recombinase, RecA protein can be bound to the homologous probe and the heterologous probe, according to the conventional method well known in the art, in the presence of co-factors. Examples of such co-factors include, but are not limited to, rATP, alone or in the presence of an rATP regeneration system, (for example, rATP regeneration system manufactured by Boehringer-Mannheim GmbH), ATP γ S, GTP γ S, dATP, a mixture of ATP γ S and ADP, a mixture of ADP and AlF_4^- (aluminium nitrate and sodium fluoride), a mixture of dADP and AlF_4^- , a mixture of ATP and AlF_4^- , and a mixture of dATP and AlF_4^- (Rigas B. et al., Proc. Natl. Acad. Sci. USA, 83: 9591-9595 (1986); Hsieh P. et al., Proc. Natl. Acad. Sci. USA, 89: 6492-6496 (1992); Ferrin L. J. et al., Science, 254: 1494-1497 (1991); Sena E. P., Nature Genetics, 3: 365-372 (1993); Teintze M. et al., Biochem. Biophys. Res. Commun., 211: 804-811 (1995); Jayasena V. K. et al., J. Mol. Biol., 230: 1015 (1993); Kowalczykowski S. C. et al., Proc. Natl. Acad. Sci. USA, 92: 3478-3482 (1995); Moreus P. L. et al., J. Biol. Chem., 264: 2302-2306 (1989); US Patent No. 4,888,274, WO93/05177, WO93/05178, WO95/18236, WO93/22443 etc.).

Preferably, co-factors are ATP (preferably, 1-5 mM), GTP γ S (preferably, 0.05-5 mM), ATP γ S (preferably, 0.01-3 mM), a mixture of ATP γ S and ADP (0.3-3 mM ATP γ S + 0.3-1.1 mM ADP), or a mixture of ADP and AlF_4^- [0.02-5 mM ADP + 0.01-0.5 mM $Al(NO_3)_3$ + 5-50 mM NaF].

The homologous probe and the heterologous probe in accordance with the present invention may be used in either a single-stranded or a double-stranded form. In general, the probe is subjected to denaturation by heating for about 5 minutes at 95-

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100°C prior to the reaction, and after chilling the probe on ice for about 20 seconds to 1 minute, the probe is used for the binding reaction with RecA-like recombinase. If desired, the probe is centrifuged for about 1-10 seconds at 0-4°C prior to the binding reaction with the recombinase. Though the denatured single-stranded probe can be
5 stored in a freezer at -20°C, preferably, it is immediately mixed with the standard reaction solution containing a co-factor and a RecA-like recombinase in an ice water bath. The single-stranded probe is bound to the RecA-like recombinase by incubating the mixed solution at 37°C for 0-20 minutes (pre-incubation is not always necessary if ATP, GTPγS, ATPγS or a mixture of ATPγS and ADP is used as a co-factor). In
10 this reaction, RecA-like recombinase generally binds to the single-stranded probe at the ratio of one molecule to about 3 nucleotides of a single-stranded probe. The binding reaction can be carried out at the mixing ratio of the amount of RecA protein (monomer) to nucleotides of the probe ranging from 5:1 to 1:8, preferably from 3:1 to 1:6. If desired, the reaction is performed in the presence of single-strand DNA binding
15 protein (SSB), topoisomeraseI or topoisomeraseII.

The homologous probe of the present invention may be used as a labeled homologous probe, when it is bound to RecA-like recombinase having various labels or ligands described in the reference WO95/18236, the text of which is incorporated
20 herein by reference. On the contrary, it is not preferable to use the heterologous probe with RecA-like recombinase having various labels or ligands.

The homologous probe/RecA-like recombinase complex and the heterologous probe/RecA-like recombinase complex are added to the DNA sample under conditions for avoiding the denaturation of the double-stranded target DNA, for example, at a temperature lower than that for melting double-stranded DNA. The mixture is
25 incubated at 37°C for generally 10 minutes-24 hours, preferably 15 minutes-2 hours to carry out homologous pairing (hybridization).

The ratio of the amount of homologous probe to that of the heterologous probe added to the reaction is generally 1:1-1:500, preferably 1:1-1:250. It is a surprise discovery of the present invention that the use of the heterologous probe together with
30 the homologous probe in a ratio as described above greatly increases the specificity of targeting, enriching, detecting and isolating of a target nucleic acid sequence. As

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described in the Examples of the present invention, for example, the method of the present invention is applied to isolate double-stranded target DNA from a sample containing about 1 µg of total DNA with a molar ratio of target DNA to non-target DNA being 1:10,000. When 200 ng of RecA protein coated homologous probe of 60-
5 275 nucleotide length is used alone, the specificity of the reaction is extremely low. A large amount of heterologous DNA other than target DNA is recovered in contamination with the double-stranded target DNA, though, a certain amount of the double-stranded target DNA is also recovered. When the amount of the RecA protein coated homologous probe is reduced to 50 ng and used alone, the efficiency of the
10 reaction decreases in comparison with the efficiency of the reaction wherein 200 ng of homologous probe is used alone, and likewise the yield of the double-stranded target DNA decreases. However, when 50 ng of the RecA protein coated homologous probe is reacted with the sample DNA in the presence of 150 ng of the RecA protein coated heterologous probe, not only the reaction efficiency is increased and a larger amount of
15 the double-stranded target DNA is recovered in comparison with the reaction wherein 200 ng of homologous probe is used alone, but also the specificity of the reaction greatly increases because of the strong inhibition of the recovery of heterologous DNA. When the amount of the homologous probe is further reduced (25 ng or 10 ng), and thus the ratio of the homologous probe to the heterologous probe is further
20 reduced (homologous probe: heterologous probe =10:190, or homologous probe : heterologous probe=25:175), the specificity of the reaction is further distinctively improved.

It should be understood that the ratio of the amount of homologous probe to that of heterologous probe, also the ratio of RecA-like recombinase bound to the
25 probes may be altered by considering the property of the sample and the like. In view of the teaching of this disclosure, one skilled in the art can alter the ratio in light of the changes in the property of the sample without undue experiments.

In an embodiment of the present invention, the reaction solution for the homologous pairing (hybridization) may be prepared so that the final concentration of
30 each component may be within the range as follows: 1-100 mM Tris-HCl buffer or Tris-acetate buffer, 1-30 mM magnesium acetate or magnesium chloride, 0-50 mM

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sodium acetate or sodium chloride, 0-3 mM dithiothreitol, 0-100 mM EGTA, 0-50 mM spermidine, 0-10% glycerol, 1-5 mM ATP or 0.05-5 mM GTP γ S or 0.01-3 mM ATP γ S or 0.3-3 mM ATP γ S + 0.3-1.1 mM ADP, 0.002-0.025 mM RecA protein, 0.5-150 ng of homologous probe and 20-200 ng of the heterologous probe per
5 reaction, and about 1 μ g of the DNA sample (target DNA + heterologous DNA).

The methods of the present invention may be used in connection with well-known methods in the art (US Patent No. 4,888,274, WO093/05177, WO093/05178, WO095/18236, Teintze M. et al., Biochem. Biophys. Res. Commun., 211: 804-811 (1995)) for detection or isolation of a complex formed between double-stranded target
10 DNA and RecA protein coated homologous probe, wherein either the double-stranded target DNA, or RecA protein, or the homologous probe may be labeled with labels or ligands. For example, the complex may be a complex of homologous probe carrying various labels or ligands/RecA protein/double-stranded target DNA, or a complex of homologous probe/RecA protein carrying various labels or ligands/double-stranded
15 target DNA, or a complex of homologous probe/ RecA protein/double-stranded target DNA carrying various labels or ligands. In accordance with the present invention, those complexes are formed by homologous pairing (hybridization reaction) using RecA-like recombinase in the presence of both a homologous probe and a heterologous probe.

20 In a preferred embodiment, a method for isolating double-stranded target DNA comprises the steps of: labeling the homologous probe with biotin; coating the homologous probe and a non-labeled heterologous probe with RecA protein in the presence of an appropriate co-factor (for example, GTP γ S or a mixture of ATP γ S and ADP); reacting the RecA coated homologous and heterologous probes with a DNA
25 sample containing double-stranded target DNA to form a complex of homologous probe/target DNA; and then capturing the complex on magnetic beads (BioMag, Dynal) conjugated with streptavidin. After washing out the DNA and probes which are not captured on the beads, biotin-labeled homologous probe/double-stranded target DNA complex bound to the beads is incubated in a solution containing NaCl at a
30 temperature between the room temperature and 85°C for about 5-15 minutes to separate (elute) the double-stranded target DNA fraction from the beads. Preferably,

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the recovered target DNA fraction is then used to transform appropriate host cells, followed by selecting the transformed cells having the target DNA, and recovering the target DNA from the transformed cells.

5 To carry out the methods described above, the present invention also provides a kit for targeting, enriching, detection and/or isolation of a double-stranded target nucleic acid sequence in a sample. The kit may comprise a carrier means being compartmentalized to receive in close confinement one or more container means such as vials, tubes, and the like, each of the container means comprising one of the separate elements to be used in the methods of the present invention.

10 For example, one of the container means may contain RecA-like recombinase. A second container may contain a solid phase designed to capture the complex of double-stranded target nucleic acid sequence and homologous probe labeled with markers or ligands. Containers are also provided for containing co-factors, heterologous probe and washing solution separately.

15 In an embodiment of the present invention, a kit in accordance with the present invention may comprise the following elements in separate compartments respectively: RecA-like recombinase, appropriate co-factors, a heterologous probe, a solid phase designed to capture the complex of double-stranded target DNA and homologous probe labeled with marker or ligand, and washing solution.

20 The method in accordance with the present invention may be used to target, enrich, detect and /or isolate with high specificity target nucleic acid molecules existing in a given sample at a very low level. In an embodiment of the present invention, the molar ratio of the target nucleic acid molecules to the non-target nucleic acid molecules may be less than 1:1,000. Preferably, the ratio may be more than
25 1:1,000,000, and more preferably more than 1:500,000.

The methods of the present invention are well suited for use in isolation and subsequent cloning of the target gene from a mixture of cDNAs or genomic DNAs as well as in screening for the target gene from various gene libraries. Examples of such gene libraries include, but are not limited to, cDNA libraries, cosmid libraries, YAC
30 libraries, and the like. Since a short strand probe may be used in the present invention,

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it becomes possible to isolate or screen a target gene whose nucleic acid sequence or amino acid sequence is only partially revealed.

The present invention may also be used in conjunction with well-known methods in the art for the following purposes: amplification of the target DNA
5 sequence using RecA-like recombinase (US Patent No 5,223,414, WO91/17267); mapping of various genes, such as RARE (RecA-Assisted Restriction Endonuclease cleavage) based mapping using an oligonucleotide probe (Ferrin L. J. et al., Nature Genetics, 6:379 (1994); Revet B. M. et al., J. Mol. Biol., 232:779 (1993)); nucleotide
10 DNA using oligonucleotide (Koob M. et al., Nucleic Acid Res., 20:5831 (1992); Golub E. I. et al., Nucleic Acid Res., 20:3121 (1992); Mikhail A. et al., Biochemistry, 34:13098 (1995)). Applications of the present invention in conjunction with the above-known methods are well within the skill in the art in view of the instant disclosure.

15 Furthermore, the present invention may be used for isolation and/or detection of the specific target DNA of interest from clinical specimens containing a mixture of cDNA or genomic DNA. Therefore, the present invention allows the diagnosis of a variety of genetic aberration or mutation, or infectious diseases caused by a variety of pathogenic microorganism or viruses.

20 Moreover, the present invention may also be used in conjunction with an in-situ hybridization method, mediated by RecA-like recombinase (WO93/05177, WO095/18236), to improve the specificity or the efficiency of the hybridization. It is also applicable as a gene therapy technique and a transgenic technique to produce transgenic animals or plants by means of gene alternation or transcription inhibition
25 using in vivo gene-targeting in the living cells (US Patent No.5,468,629, WO093/2244; Golub E. I. et al., Nucleic Acid Res., 20: 3121 (1992); Golub E.I. et al., Proc. Natl. Acad. Sci. USA, 90; 7186 (1993), etc.). Applications of the present invention in conjunction with the above known methods are well within the skill in the art in view of the instant disclosure.

30 For example, when the present invention is used for in vivo gene-targeting in living cells, both RecA-like recombinase coated homologous probe and heterologous

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probe are used in forming the complex of RecA-like recombinase mediated homologous probe and double-stranded target DNA. Accordingly, a method for targeting a double-stranded nucleic acid target sequence in a sample of living cells by in vivo gene targeting method using a RecA-like recombinase may comprise the following steps: (a) providing a recombinase coated homologous probe and a recombinase coated heterologous probe; (b) introducing the homologous probe and the heterologous probe into the living cells; and (c) incubating said living cells for a sufficient period of time to allow the homologous probe to be transformed into the genome of the cells.

The conventional method of in vivo gene-targeting in living cells, which may be used in conjunction with the present invention, is described in US Patent No.5,468,629 and WO093/2244, the relevant text of which is incorporated herein in its entirety by reference.

The following are examples provided to describe the present invention. They are intended only for illustrating the present invention and not as limiting the scope of the present invention.

EXAMPLE I

Preparation of target DNA, heterologous DNA, homologous probe and heterologous probe

1. Preparation of the double-stranded target DNA and the heterologous DNA
Php53B, a 6.6 kb plasmid containing a complete sequence of cDNA corresponding to the sequence of human tumor suppressor gene p53 (Zakut-Houri R. et al., EMBO J., 4: 1251 (1985)) was used as double-stranded target DNA in a circular form, and plasmid vector pUC18 (2.7 kb) was used as heterologous double-stranded DNA in circular form. These plasmids were purified from the cells of *Escherichia coli*, accommodating each plasmid, using QIAGEN Plasmid Maxi Kit (manufactured by QIAGEN GmbH).

2. Preparation of the homologous probe

The double-stranded DNA fragments of 275bp (SEQ ID NO:1) and 60bp (SEQ ID NO:2), which contain the partial sequence of p53 cDNA, were prepared

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under the standard amplification condition (PCR method) using Taq polymerase and the following primers: 5'-CCTTGCCGTCCCAAGC AATGGATGA-3' (SEQ ID NO:11, and corresponding to the 1-25th nucleotide sequence of the SEQ ID NO:1), 5'-CGTGCAAGTCACAGACTTGGCTGTC-3' (SEQ ID NO:12, and corresponding to the 251-275th nucleotide sequence of the SEQ ID NO:1), 5'-AGCTACGGTTTCCGTCTGGGCTTCT-3' (SEQ ID NO:13, and corresponding to the 1-25th nucleotide sequence of the SEQ ID NO:2), 5'-CGTGCAAGTCACAGACTTGGCTGTC-3' (SEQ ID NO:12, and corresponding to the 36-60th nucleotide sequence. of SEQ ID NO:2). The above primers are biotinylated at their 5'-ends. The double-stranded fragments were used as homologous probes after being denatured into single-stranded DNA by heating immediately before use. The nucleotide sequences corresponding to these regions of p53 cDNA are shown as SEQ ID NOS:1 and 2.

The double-stranded DNA fragments of 410 bp (SEQ ID NO:9), 779 bp (SEQ ID NO:10), and 1,317 bp (SEQ ID NO:8), which contain the partial sequence of p53 cDNA, were also prepared under standard amplification conditions (PCR method) using Taq polymerase and the following primers (manufactured by GENOSYS): 5'-CCTTGCCGTCCCAAGCAATGGATGA-3' (SEQ ID NO:11, and corresponding to the 236-260th nucleotide sequence of the SEQ ID NO:8), 5'-CGTCATGTGCTGTGACTGCTTGTAG-3' (SEQ ID NO:14, and corresponding to the 621-645th nucleotide sequence of the SEQ ID NO:8), 5'-CCTTGCCGTCCCAAGCAATGGATGA-3' (SEQ ID NO:11, and corresponding to the 236-260th nucleotide sequence of the SEQ ID NO:8), 5'-CCCTTTCTTGCGGAGATTCTCTTCC-3' (SEQ ID NO:15, and corresponding to the 990-1,014th nucleotide sequence. of SEQ ID NO:8), 5'-GTCTAGAGCCACCGTCCAGGGAGCA-3' (SEQ ID NO:16, and corresponding to the 1- 25th nucleotide sequence of SEQ ID NO:8), and 5'-TCAGTCTGAGTCAGGCCCTTCTGTC-3' (SEQ ID NO:17, and corresponding to the 1,293-1,317th nucleotide sequence of SEQ ID NO:8). The above primers are biotinylated at their 5'-ends. These double-stranded fragments were used as homologous probe after denatured into single-stranded DNA by heating immediately

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before the use. The nucleotide sequences corresponding to these regions of p53 cDNA are shown as SEQ ID NOS: 9, 10 and 8, respectively.

3. Preparation of the heterologous probe

The double-stranded DNA fragments of 500 bp (SEQ ID NO: 3) and 1,393 bp (SEQ ID NO:4), which contain a partial sequence of the genome of lambda phage, were prepared under standard amplification conditions (PCR method) using Taq polymerase and the following primers: 5'-GATGAGTTCG TGTCCGTACAACTGG-3' (SEQ ID NO:18, and corresponding to the 1-25th nucleotide sequence of SEQ ID NO:3), 5'GGTTATC GAAATCAGCC ACAGCGCC-3' (SEQ ID NO:19, and corresponding to the 476-500th nucleotide sequence of SEQ ID NO:3), 5'-GCGGCACGGAGTGGAGCAAGCGTGA-3' (SEQ ID NO:20, and corresponding to the 1-25th nucleotide sequence of SEQ ID NO:4), 5'-ATACGGCTGAGGTTTTCAACGGCCT-3' (SEQ ID NO:21, and corresponding to the 1369-1393th nucleotide sequence of SEQ ID NO:4). The double-stranded fragments were used as heterologous probes after denatured into single-stranded DNA by heating immediately before the use. The nucleotide sequences corresponding to these regions of lambda phage are shown as SEQ ID NOS:3 and 4.

Also, all lambda phage DNA (manufactured by Nippon Gene) are fragmented under the nick-translation reaction in the presence of dNTPs using a commercially available nick-translation kit (manufactured by BRL). The fragments were used as heterologous probes after being denatured into single-stranded DNA by heating immediately before use. ("Lambda DNA fragments" are formed from fragments with 100-1,500 nucleic acid in length).

Salmon sperm DNA (Type III, Sigma) was used as a heterologous probe after it was fragmented and heat-denatured according to conventional methods (Maniatis T. et al., "Molecular Cloning") and then denatured into single-stranded DNA again immediately before the use. Fragments of Salmon sperm DNA have a length of about 100 to 1,500 nucleotides.

Commercially available single strand DNA of M13mp18 (manufactured by New England Biolabs Inc.) was purchased and also used as the heterologous probe.

EXAMPLE 2

Isolation of double-stranded target DNA using RecA protein

1. Target DNA: heterologous DNA = 1 : 10,000 (when GTP γ S was used as a co-factor)

5 A. Homologous pairing (hybridization reaction)

(a) preparation of various probe/ RecA protein complexes

The double-stranded homologous probes prepared in EXAMPLE 1 2, such as 275bp, 60bp, 410bp, 779bp, or 1,317bp sequence, or the mixture of the homologous probes and the various heterologous probes prepared in EXAMPLE 1 3, were diluted
10 in either distilled water or TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 7.5). The aliquot was transferred to a 0.6 ml microcentrifuge tube and was heated in boiling water for 5 minutes to denature the double-stranded probe. The tube was then rapidly chilled in ice water. 0.9 μ l of 10 x reaction buffer, [300 mM Tris-HCl (pH 7.5 at 37°C), 20 mM MgCl₂, 4 mM DTT, 30% glycerol], 0.9 μ l of 3 mM GTP γ S solution,
15 and RecA protein (manufactured by Boehringer-Mannheim GmbH) were added to the tube to form a mixture. The mixture was diluted with distilled water to a total volume of 9 μ l and was allowed to react for 12 minutes at 37°C to allow the formation of probe/RecA complex.

(b) Preparation of probe/target DNA complex

20 Then 0.9 μ l of 10x reaction buffer, 0.9 μ l of 3 mM GTP γ S solution, 0.9 μ l of 0.2 M MgCl₂ solution, 245 pg. of php53B, which was double-stranded target DNA in circular form prepared as in EXAMPLE 1. 1, and 1 μ g of pUC18, which was circular heterologous DNA (molar ratio of target DNA to heterologous DNA is 1 : 10,000) were mixed and diluted with distilled water to a total volume of 9 μ l. This mixture was
25 mixed with either the complex of homologous probe/ RecA protein or the mixture of homologous probe/ RecA protein complex and heterologous probe/ RecA protein complex obtained from the reaction described above. The mixture thus obtained was subjected to the reaction for 60 minutes at 37°C to allow the hybridization and formation of the complex of probe/target DNA. The type and the amount of the
30 homologous probe, those of the heterologous probe and the amount of RecA protein

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which were used in each reaction are shown in Table 1. In all reactions, a final volume of the reactant was 18 μ l.

B. Capturing/isolation by magnetic beads

20 μ l of the magnetic beads coated with straptavidin, manufactured by
5 DYNAL®, was transferred to a 0.6 ml microcentrifuge tube and was washed two times in 100 μ l of 33 mM Tris-HCl and 50 mM NaCl (pH7.5) using a Magnetic Beads Separation Rack (MAGNA-SEPTM). After discarding the washing solution, the washed magnetic beads were mixed with the whole volume of the hybridization reaction solution obtained above in the 0.6 ml microcentrifuge tube, and this mixture
10 was mixed well and stored for 15 minutes at room temperature with stirring every 2-3 minutes. The magnetic beads were then separated from supernatant using the Magnetic Beads Separation Rack. After discarding the supernatant, magnetic beads were washed twice in 100 μ l of 33 mM Tris-HCl and 50 mM NaCl (pH7.5) solution for 5 minutes at 37°C to get rid of any unbound molecules such as probes, RecA
15 proteins, target DNA and heterologous DNA molecules and the like. After discarding the washing solution, the beads were suspended and mixed with 10 μ l of 33 mM Tris-HCl and 200 mM NaCl (pH7.5), and then the mixture was heated to 85°C for 8 minutes. The supernatant containing target DNA was recovered using the Magnetic Beads Separation Rack.

20 C. Transformation

100 μ l of competent cells, which were prepared from *Escherichia coli* strain JM109 according to the method of Nojima et al. (Inoue H. et al., Gene, 96:23 (1990)), was transferred into a 1.5ml microcentrifuge tube, and mixed with 2 μ l of the supernatant removed from 10 μ l of supernatant that recovered in B. This mixture was
25 then stored for 30 minutes on ice. The mixture was subsequently incubated for 30 seconds at 42°C, and then chilled in the ice again for 1-2 minutes. Then 0.5 ml of SOC medium (2% Bacto trypton, 0.5% Bacto yeast extract, 10 mM NaCl, 2.5 mM KCl, 10 mM MgCl₂, 10 mM MgSO₄, 20 mM glucose) was added to the tube and incubated at 37°C for 1 hour with shaking. The cells were spread on LB Plate,
30 containing 50-100 μ g/ml ampicillin, 1 mg/plate X-gal (5-bromo-4-chloro-3-indolyl- β - D-galactoside), 0.5-1.0 mg/plate IPTG (isopropyl- β -D(-)-thiogalactopyranoside), and

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incubated over night at 37° C. Under this condition, *Escherichia coli* transformed with php53B, the double-stranded target DNA, shows white colonies, and *Escherichia coli* transformed with pUC18, the heterologous DNA, shows blue colonies.

The various conditions and the results of the transformation obtained under
5 those conditions are shown in Tables 1 and 2.

Table 1

| Reaction Condition | | | | | | |
|--------------------|-----|-----------------|-------------------|----------------------|---------------------|----------------|
| | No. | Homo.P. Type | Homo.P. Amount | Hetero. P. Type | Hetero.P. Amount | RecA Amount |
| 10 | 1 | 1,317 bases | 200ng | - | - | 6.0µg |
| | 2 | 1,317 bases | 67ng | lambda DNA fragments | 133ng | 6.0µg |
| | 3 | 779 bases | 200ng | - | - | 6.0µg |
| | 4 | 779 bases | 100ng | lambda DNA fragments | 100ng | 6.0µg |
| | 5 | 779 bases | 67ng | lambda DNA fragments | 133ng | 6.0µg |
| 15 | 6 | 779 bases | 20ng | lambda DNA fragments | 180ng | 6.0µg |
| | 7 | 410 bases | 200ng | - | - | 6.0µg |
| | 8 | 410 bases | 100ng | lambda DNA fragments | 100ng | 6.0µg |
| | 9 | 410 bases | 67ng | lambda DNA fragments | 133ng | 6.0µg |
| | 10 | 410 bases | 10ng | lambda DNA fragments | 190ng | 6.0µg |
| 20 | 11 | 275 bases | 200ng | - | - | 6.0µg |
| | 12 | 275 bases | 50ng | - | - | 6.0µg |
| | 13 | 275 bases | 50ng | - | - | 1.5µg |
| | 14 | 275 bases | 133ng | lambda 500 bases | 67ng | 6.0µg |
| | 15 | 275 bases | 100ng | lambda 500 bases | 100ng | 6.0µg |
| 25 | 16 | 275 bases | 67ng | lambda 500 bases | 133ng | 6.0µg |
| | 17 | 275 bases | 50ng | lambda 500 bases | 150ng | 6.0µg |
| | 18 | 275 bases | 10ng | lambda 500 bases | 190ng | 6.0µg |
| | 19 | 275 bases | 10ng | lambda 1400 bases | 190ng | 6.0µg |
| | 20 | 275 bases | 10ng | salmon sperm DNA | 190ng | 6.0µg |
| 30 | 21 | 275 bases | 10ng | M13mp18 | 190ng | 6.0µg |

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| | | | | | | |
|----|----|-----------|-------|----------------------|-------|--------|
| | 22 | 275 bases | 1ng | lambda DNA fragments | 250ng | 7.5µg |
| | 23 | 275 bases | 1ng | lambda DNA fragments | 500ng | 15.0µg |
| | 24 | 60 bases | 200ng | - | - | 6.0µg |
| | 25 | 60 bases | 50ng | - | - | 6.0µg |
| 5 | 26 | 60 bases | 50ng | - | - | 1.5µg |
| | 27 | 60 bases | 133ng | lambda 500 bases | 67ng | 6.0µg |
| | 28 | 60 bases | 100ng | lambda 500 bases | 100ng | 6.0µg |
| | 29 | 60 bases | 67ng | lambda 500 bases | 133ng | 6.0µg |
| | 30 | 60 bases | 50ng | lambda 500 bases | 150ng | 6.0µg |
| 10 | 31 | 60 bases | 25ng | lambda 500 bases | 175ng | 6.0µg |
| | 32 | 60 bases | 25ng | lambda 1400 bases | 175ng | 6.0µg |
| | 33 | 60 bases | 25ng | salmon sperm DNA | 175ng | 6.0µg |
| | 34 | 60 bases | 25ng | M13mpl8 | 175ng | 6.0µg |
| | 35 | 60 bases | 10ng | lambda 500 bases | 190ng | 6.0µg |

15 As used herein, "homo.p." is a homologous probe; "hetero.p." is a heterologous probe.

All the "lambda DNA fragments" used in table 1 are prepared by a nick-translation of whole lambda phage DNA in the presence of dNTPs using an on-the-shelf nick-translation kit (manufactured by BRL Co., Ltd.)

Table 2

20 The Results of Transformation¹⁾

| | Reaction No. | Number of white colonies | Number of blue colonies | specificity (%) ²⁾ |
|----|--------------|--------------------------|-------------------------|-------------------------------|
| | 1 | 472 | 20,458 | 2.3 |
| | 2 | 385 | 861 | 30.9 |
| 25 | 3 | 590 | 18,760 | 3.0 |
| | 4 | 601 | 1,719 | 25.9 |
| | 5 | 499 | 807 | 38.2 |
| | 6 | 461 | 330 | 58.3 |
| | 7 | 904 | 18,362 | 4.7 |

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| | | | | |
|----|----|-------|--------|------|
| | 8 | 1,096 | 4,123 | 21.0 |
| | 9 | 1,118 | 2,678 | 33.8 |
| | 10 | 756 | 252 | 75.0 |
| | 11 | 830 | 22,238 | 3.6 |
| 5 | 12 | 756 | 11,340 | 6.3 |
| | 13 | 406 | 13,036 | 3.0 |
| | 14 | 1,540 | 10,380 | 12.9 |
| | 15 | 1,626 | 2,254 | 41.9 |
| | 16 | 1,488 | 2,357 | 38.7 |
| 10 | 17 | 1,744 | 2,688 | 39.4 |
| | 18 | 1,224 | 336 | 78.5 |
| | 19 | 1,046 | 330 | 76.0 |
| | 20 | 1,024 | 270 | 79.1 |
| | 21 | 938 | 254 | 78.7 |
| 15 | 22 | 543 | 146 | 78.8 |
| | 23 | 302 | 199 | 60.3 |
| | 24 | 126 | 3,264 | 3.7 |
| | 25 | 88 | 3,046 | 2.8 |
| | 26 | 6 | 1,020 | 0.6 |
| 20 | 27 | 610 | 1,030 | 37.2 |
| | 28 | 605 | 786 | 43.5 |
| | 29 | 622 | 650 | 48.9 |
| | 30 | 604 | 616 | 49.5 |
| | 31 | 606 | 270 | 69.2 |
| 25 | 32 | 484 | 244 | 66.5 |
| | 33 | 646 | 266 | 70.8 |
| | 34 | 330 | 320 | 50.8 |
| | 35 | 502 | 222 | 69.3 |

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- 1): Mean values obtained from the three repeated experiments are indicated.
- 2): $\text{Specificity (\%)} = \frac{\text{number of white colonies}}{\text{number of white colonies} + \text{number of blue colonies}} \times 100$

The above results show that when a large amount (200 ng) of a homologous probe is used, the reaction specificity is extremely low, although the reaction efficiency is relatively high. As used herein, the reaction specificity is determined based on the number of white colonies versus the total number of white and blue colonies. The reaction efficiency is determined based on the number of white colonies and the amount of target DNA recovered.

For example, when a large amount (200ng) of a single species of homologous probe, which is longer than 275 nucleotides, was used (see Nos. 1, 3, 7, 11), the reaction efficiency was considered relatively high, since the large number of *Escherichia coli* colonies containing target DNA (the number of white colonies) were observed, and the yield of target DNA was also high. At the same time, however, the number of *Escherichia coli* colonies containing the heterologous DNA (blue colonies) greatly exceeded the number of white colonies, which indicated the reaction specificity was considered to be extremely low.

On the other hand, when a large amount (200ng) of single species of homologous probe of 60 nucleotide long was used (No.24), not only the specificity of the reaction was extremely low, but also the efficiency of the reaction was significantly lower. The specificity of the reaction is low due to the high level of contamination of the heterologous DNA. The efficiency of the reaction is lower since the yield of the target DNA was greatly reduced in comparison with the yield of the target DNA when the homologous probe of 275 nucleotide was used. When the amount of the homologous probe or RecA protein was reduced (Nos. 12, 13, 25, 26), the contamination of the heterologous probe was slightly inhibited, and the yield of the target DNA also decreased. Subsequently, both the reaction efficiency and the specificity were greatly reduced.

However, when a small amount of the homologous probe (50ng) was used with a large amount (150ng) of the heterologous probe (No. 17, 30) in the reaction, the

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specificity of the reaction was improved. This is due to the inhibition of contamination of the heterologous DNA. When the amount of the homologous probe was further reduced (10ng of the homologous probe of 275 nucleotides in length, 25ng or 10ng of homologous probe of 60 nucleotides in length), and the amount of the heterologous probe was further increased (175ng or 190ng) (Nos. 18-21, 31-35), the level of contamination of the heterologous DNA was dramatically reduced in comparison to Nos. 17 and 30, and thus the specificity of the reaction was remarkably improved.

When a reduced amount of the homologous probe (10-133 ng) was used with a large amount (67-190 ng) of the heterologous probe in the reaction (Nos. 8, 9, 14-21, 27-35), the yield of the target DNA was dramatically increased, and therefore, the reaction efficiency itself was considered to be improved. When a short homologous probe such as a probe of 60-410 nucleotide long (the length of a complementary region between target DNA and probe is short) was used in combination with various species of heterologous probes, it was shown that not only the reduction of the reaction efficiency, resulting from using a reduced amount of homologous probe alone, was recovered, but also the reaction efficiency was improved when compared to the efficiency achieved by using an unreduced amount of homologous probe alone.

The above results show that when the heterologous probe that is 1-500 fold of the amount of homologous probe is used (Nos. 2, 4-6, 8-10, 15-23, 28-35), the specificity of the reaction is substantially improved. The specificity is improved more than four times as compared with those obtained when single species of the homologous probe is used without adding the heterologous probe.

2. Target DNA: heterologous DNA = 1: 100,000 (GTPyS was used as a co-factor)

A. Preparation of the heterologous probe

In addition to the 500bp fragments of the double-stranded DNA prepared in EXAMPLE 1. 3, containing the partial sequence of the genome of lambda phage and salmon sperm DNA, 308 bp of the double-stranded fragments (SEQ ID NO:5), containing the partial sequence of lambda phage, was newly prepared under standard

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amplification conditions (PCR method) using a primer, 5'-AGGTGCGGTGATACGTGGTGTTCCTT -3' (SEQ ID NO:22, and corresponding to the 1-25th nucleotide sequence of SEQ ID NO: 5), another primer, 5'-ATACGGCTGAGGTTTCAACGGCCT -3' (SEQ ID NO:21, and corresponding to the 284-308th nucleotide sequence of SEQ ID NO: 5), and Taq polymerase. This fragment was used as a heterologous probe after being denatured into single-stranded DNA by heating immediately before the use. The DNA sequence of the fragment is set forth in SEQ ID NO:5.

B. Homologous pairing (hybridization reaction)

10 (a) the preparation of various probe/ RecA protein complex

The double-stranded homologous probe consisting of 275bp or 60bp sequence prepared in EXAMPLE 1. 2, containing a partial sequence of p53 cDNA and biotinylated at its 5'-end, or the mixture of the said homologous probe and the various heterologous probes described in EXAMPLE 2. 2. A was diluted in either distilled water or TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 7.5). The aliquot was transferred to a 0.6 ml microcentrifuge tube and was heated in the boiling water for 5 min to denature the double-stranded probe. The tube was rapidly chilled in ice water, then 0.9 μ l of 10 x reaction buffer [300 mM Tris-HCl (pH 7.5 at 37°C), 20 mM MgCl₂, 4 mM DTT, 30% glycerol], 0.9 μ l of 5 mM GTP γ S solution, and RecA protein (manufactured by Boehringer-Mannheim GmbH) were added to the tube. The mixture was allowed to react for 12 minutes at 37°C after dilution with distilled water to a total volume of 9 μ l

(b) The preparation of probe/target DNA complex

Then 0.9 μ l of 10x reaction buffer, 0.9 μ l of 5 mM GTP γ S solution, 0.9 μ l of 0.2 M MgCl₂ solution, 24.5 pg of php53B which was double-stranded target DNA in circular form prepared as in EXAMPLE 1. 1, and 1 μ g of pUC18 which was circular heterologous DNA (molar ratio of target DNA to heterologous DNA is 1 : 100,000) were mixed and diluted with distilled water to a total volume of 9 μ l. This mixture was mixed with either the complex consisting of homologous probe and RecA protein or the mixture of homologous probe/ RecA protein complex and heterologous probe/ RecA protein complex obtained from the reaction described above, and the mixture

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thus obtained was subjected to the reaction for 60 minutes at 37°C. The type and the amount of the homologous probe, those of the heterologous probe and the amount of RecA protein which were used in each reaction are shown in Table 3. In all the reactions, a final volume of the reactant was 18 μ l.

5 C. Capturing/isolation by magnetic beads

20 μ l of the magnetic beads coated with streptavidin, manufactured by DYNAL®, were transferred to a 0.6 ml microcentrifuge tube and were washed two times in 100 μ l of 33 mM Tris-HCl and 50 mM NaCl (pH7.5) using Magnetic Beads Separation Rack (MAGNA-SEPTM). After discarding the washing solution, the
10 washed magnetic beads were mixed with the whole volume of the hybridization-reaction solution obtained above in the 0.6 ml microcentrifuge tube, and this mixture was mixed well and stored for 15 minutes at room temperature with stirring every 2-3 minutes. The magnetic beads were then separated from the supernatant using the Magnetic Beads Separation Rack. After discarding the supernatant, the magnetic
15 beads were washed twice in 100 μ l of 33 mM Tris-HCl and 50 mM NaCl (pH7.5) solution for 5 minutes at 37°C to get rid of any unbounded molecules such as probes, RecA proteins, target DNA, heterologous DNA etc. After discarding the washing solution, the beads were suspended and mixed with 10 μ l of 33 mM Tris-HCl and 200 mM NaCl (pH7.5), and then the mixture was heated to 85°C for 8 minutes. The
20 supernatant containing target DNA was recovered using Magnetic Beads Separation Rack.

 D. Transformation

100 μ l of competent cells, which were prepared from *Escherichia coli* strain JM109 according to the method of Nojima et al. (Inoue H. et al., Gene, 96:23 (1990)),
25 were transferred into a 1.5ml microcentrifuge tube, and mixed with 10 μ l of the supernatant recovered in EXAMPLE 2.2.C. This mixture was then stored for 30 minutes on ice. The mixture was subsequently incubated for 30 seconds at 42°C, and then chilled in ice again for 1-2 minutes. 0.5 ml of SOC medium (2% Bacto trypton, 0.5% Bacto yeast extract, 10 mM NaCl, 2.5 mM KCl, 10 mM MgCl₂, 10 mM
30 MgSO₄, 20 mM glucose) was added to the tube and incubated at 37°C for 1 hour with shaking. The cells were spread on LB Plate [containing 50-100 μ g/ml ampicillin, 1

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mg/plate X-gal (5-bromo-4-chloro-3-indolyl- β -D-galactoside), 0.5-1.0 mg/plate IPTG (isopropyl- β -D(-)-thiogalactopyranoside)] and incubated over night at 37°C. Under this condition, *Escherichia coli* cells transformed with php53B, the double-stranded target DNA, show white colonies, and *Escherichia coli* cells transformed with pUC18, heterologous DNA, show blue colonies.

The various conditions and the results of the transformation obtained under those conditions are shown in Table 3 and Table 4.

Table 3

| 10 | Reaction Condition | | | | | |
|----|--------------------|-----------------|-------------------|--------------------|---------------------|----------------|
| | No. | Homo.P. Type | Homo.P. Amount | Hetero. P. Type | Hetero.P. Amount | RecA Amount |
| | 1 | 275 bases | 150ng | - | - | 4.5 μ g |
| | 2 | 275 bases | 1ng | lambda 500 bases | 150ng | 4.5 μ g |
| | 3 | 275 bases | 1ng | lambda 308 bases | 150ng | 4.5 μ g |
| 15 | 4 | 275 bases | 1ng | salmon sperm DNA | 150ng | 4.5 μ g |
| | 5 | 60 bases | 150 ng | - | - | 4.5 μ g |
| | 6 | 60 bases | 1ng | lambda 500 bases | 150ng | 4.5 μ g |
| | 7 | 60 bases | 1ng | lambda 308 bases | 150ng | 4.5 μ g |
| | 8 | 60 bases | 1ng | salmon sperm DNA | 150ng | 4.5 μ g |

Table 4

| The Result of Transformation ¹⁾ | | | | |
|--|---------------------------|-----------------------------|----------------------------|----------------------------------|
| | Reaction Condition No. | Number of white colonies | Number of blue colonies | Specificity (%) ²⁾ |
| | 1 | 838 | 44,672 | 1.8 |
| 25 | 2 | 762 | 263 | 74.3 |
| | 3 | 813 | 230 | 77.9 |
| | 4 | 844 | 237 | 78.1 |
| | 5 | 13 | 6,558 | 0.2 |
| | 6 | 644 | 211 | 75.3 |

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| | | | |
|---|-----|-----|------|
| 7 | 676 | 202 | 77.0 |
| 8 | 688 | 181 | 79.2 |

1): mean values obtained from the three repeated experiments are indicated.

2): specificity (%) = number of white colonies ÷ (number of white colonies and number
5 of blue colonies) x 100

The results summarized above show that, in a reaction containing the circular target DNA at the ratio of 1:100,000, although the reaction contained an extremely small amount of the homologous probe (1 ng) and 150 fold of the amount (150ng) of the heterologous probe, the reduction in the reactivity was not observed. On the
10 contrary, the results show that a remarkable enhancement of the specificity and the reactivity by adding the heterologous probe (especially when the 60 nucleotide probe was employed). The results achieved here are the same as the results achieved in Example 2.1 when the target DNA is present at a ratio of 1:10,000. The above results also show that the 308-nucleotide probe which has a partial sequence of lambda phage
15 genome can be used as a heterologous probe.

3. Target DNA: heterologous DNA = 1 : 10,000 (a probe of 30 nucleotides in length was used and GTPγS was used as a co-factor)

A. The Preparation of the Homologous Probe

The 30 nucleotide single-stranded probe (SEQ ID NO:6) containing the partial
20 sequence of p53 cDNA was synthesized using a DNA synthesizer. The nucleotide probe was biotinylated at its 5'-end. The nucleotide sequence corresponding to this region of p53 cDNA is shown as SEQ ID NO:6.

B. The Preparation of the Heterologous Probe

In addition to the heterologous probe used in EXAMPLE 2.2, a single-stranded
25 probe of 60 nucleotides in length (SEQ ID NO:7), which contained a partial sequence of lambda phage genome, synthesized according to conventional methods using DNA synthesizer (the nucleotide sequence of this region is indicated as SEQ ID NO: 7), and rRNA from *E.coli* (a mixture of 16S and 23S, manufactured by Boehringer-Mannheim GmbH) were also used as heterologous probes.

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C. Homologous Pairing (Hybridization Reaction)

(a) The preparation of various probe/RecA protein complex

A single-stranded probe of 30 nucleotides in length containing a partial sequence of p53 cDNA and biotinylated at its 5'-end, which was prepared as in
5 EXAMPLE 2.3.A, or a mixture of the said homologous probe and various heterologous probes described in EXAMPLE 2.3.B, was diluted in either distilled water or TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 7.5). The aliquot was transferred to a 0.6 ml microcentrifuge tube and was heated in the boiling water for 5 minutes to denature the double-stranded probe. The tube was rapidly chilled in ice
10 water, then 0.9 μ l of 10 x reaction buffer [300 mM Tris-HCl (pH 7.5 at 37°C), 20 mM MgCl₂, 4 mM DTT, 30% glycerol], 0.9 μ l of 5 mM GTP γ S solution, and RecA protein (manufactured by Boehringer-Mannheim GmbH) were added to the tube, and the mixture was allowed to react for 12 minutes at 37°C after dilution with distilled water to the total volume of 9 μ l.

15 (b) The preparation of probe/target DNA complex

0.9 μ l of 10x reaction buffer, 0.9 μ l of 5 mM GTP γ S solution, 0.9 μ l of 0.2 M MgCl₂ solution, 245 pg. of php53B, which was double-stranded target DNA in circular form prepared in EXAMPLE 1 1, and 1 μ g of pUC18, which was circular
20 heterologous DNA (molar ratio of target DNA to heterologous DNA is 1: 10,000) were mixed and diluted with distilled water to a total volume of 9 μ l. This mixture was mixed with either the complex of homologous probe and RecA protein or the mixture of homologous probe/ RecA protein complex and heterologous probe/ RecA protein complex obtained from the reaction described above. The mixture thus
25 obtained was subjected to the reaction for 60 minutes at 37°C. The amount of the homologous probe, the type and the amount of the heterologous probe, and the amount of RecA protein which were used in each reaction are shown in Table 5. In all the reactions, a final volume of the reactant was 18 μ l.

D. Capturing/isolation by magnetic beads

20 μ l of the magnetic beads coated with streptavidin, manufactured by
30 DYNAL[®], was transferred to a 0.6 ml microcentrifuge tube and was washed two times in 100 μ l of 33 mM Tris-HCl and 50 mM NaCl (pH7.5) using Magnetic Beads

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Separation Rack (MAGNA-SEP™). After discarding the washing solution, whole volume of the hybridization-reaction solution was added to the microcentrifuge tube to be well mixed with the washed magnetic beads. The mixture was stored for 15 minutes at room temperature with stirring every 2-3 minutes. The magnetic beads were separated from the non-binding molecules using a Magnetic Beads Separation Rack. After discarding the supernatant, magnetic beads were washed twice in 100 µl of 33 mM Tris-HCl and 50 mM NaCl (pH7.5) solution for 5 minutes at 37°C. After discarding the washing solution, the beads were suspended and mixed with 10 µl of 33 mM Tris-HCl and 200 mM NaCl (pH7.5), and then the mixture was heated to 85°C for 8 minutes. The supernatant containing target DNA was recovered using the Magnetic Beads Separation Rack.

E. Transformation

100 µl of competent cells, which were prepared from *Escherichia coli* strain JM109 according to the method of Nojima et al. (Inoue H. et al., Gene, 96:23 (1990)), were transferred into a 1.5ml microcentrifuge tube, and mixed with 2 µl of the supernatant recovered in D. This mixture was then stored for 30 minutes on ice. The mixture was subsequently incubated for 30 seconds at 42°C, and then chilled in the ice again for 1-2 minutes. 0.5 ml of SOC medium (2% Bacto trypton, 0.5% Bacto yeast extract, 10 mM NaCl, 2.5 mM KCl, 10 mM MgCl₂, 10 mM MgSO₄, 20 mM glucose) was added to the tube and incubated at 37°C for 1 hour with shaking. The cells were spread on LB Plate [containing 50-100 µg/ml ampicillin, 1 mg/plate X-gal (5-bromo-4-chloro-3-indolyl-β-D galactoside), 0.5-1.0 mg/plate IPTG (isopropyl-β-D(-)-thiogalactopyranoside)], and incubated over night at 37°C. Under this condition, *Escherichia coli* transformed with php53B, the double-stranded target DNA, shows white colonies, and *Escherichia coli* transformed with pUC18, heterologous DNA, shows blue colonies. The various conditions and the results of the transformation obtained under those conditions are shown in Table 5 and Table 6.

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Table 5

| Reaction Condition | | | | | |
|--------------------|-----|-------------------|--------------------|---------------------|----------------|
| | No. | Homo.P. Amount | Hetero. P. Type | Hetero.P. Amount | RecA Amount |
| 5 | 1 | 150ng | - | - | 4.5μg |
| | 2 | 25ng | lambda 500 bases | 125ng | 4.5μg |
| | 3 | 25ng | lambda 308 bases | 125ng | 4.5μg |
| | 4 | 25ng | lambda 60 bases | 125ng | 4.5μg |
| | 5 | 25ng | salmon sperm DNA | 125ng | 4.5μg |
| 10 | 6 | 25ng | E.coli rRNA | 125ng | 4.5μg |

Table 6The Result of Transformation¹⁾

| | Reaction Condition | Number of white colonies | Number of blue colonies | Specificity (%) ²⁾ |
|----|-----------------------|-----------------------------|----------------------------|----------------------------------|
| 15 | 1 | 89 | 901 | 9.0 |
| | 2 | 545 | 276 | 66.4 |
| | 3 | 563 | 289 | 66.1 |
| | 4 | 596 | 280 | 68.0 |
| | 5 | 632 | 271 | 70.0 |
| 20 | 6 | 551 | 388 | 58.7 |

1): mean values obtained from the three repeated experiments are indicated.

2): specificity (%) = number of white colonies / (number of white colonies and number of blue colonies) x 100

The above results show that the specificity and the reactivity were remarkably enhanced by adding heterologous probe to the reaction when the 30 nucleotide homologous probe, as well as the 60-nucleotide and the 275-nucleotide homologous probe, were used. The results also show that the 60-nucleotide single-stranded DNA probe, containing the partial sequence of lambda phage genome, and rRNA derived

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from *Escherichia coli* (a mixture of 16S and 23 S) can be used as the heterologous probe.

EXAMPLE 3

SCREENING OF TARGET cDNA FROM cDNA LIBRARY

5 Plasmids containing p53cDNA sequence were obtained from 5µg of a cDNA library prepared from a commercially available human brain cDNA library (SUPER SCRIPT Human Brain cDNA Library manufactured by BRL). The ratio of p53 cDNA to the library DNA was determined by the colony hybridization reaction method and was about 1:100,000-1:200,000.

10 The double-stranded homologous probe of 275bp and 60bp sequence, and the single-stranded homologous probe of 30 bp, as prepared in EXAMPLE 1 and EXAMPLE 2, were used. The lambda DNA fragments, as prepared in EXAMPLE 1, were used as heterologous probe.

 Homologous pairing, capturing/isolation by magnetic beads and transformation
15 were performed in accordance with the methods described in EXAMPLE 2.2 and EXAMPLE 2.3. The obtained transformants were spread on LB Plate containing 50-100 µg/ml ampicillin, and incubated over night at 37°. 30 colonies were randomly selected from the ampicillin-resistant colonies obtained by each of the reactions, and plasmid DNA was extracted from each of the colonies by the Miniprep. method.
20 (Quantum Prep® Plasmid Miniprep Kit manufactured by Bio Rad was used.)

 After performing the PCR reaction using a primer that has the same sequence as the one used in the preparation of the 275bp homologous probe and that is not biotinylated, agarose electrophoretic analysis was performed to determine, based on the presence of the 275 nucleotide fragments, whether or not the plasmid DNA
25 extracted from each colony contained p53 cDNA. The specificity was calculated based on the following formula:

$$\text{Specificity (\%)} = \text{Number of PCR positive clones} \div 30 \times 100$$

 The various reaction conditions and the cDNA library screening results obtained under those reaction conditions are shown in Table 7 and 8.

30

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Table 7

| No. | Homo.P. Type | Homo.P. Amount | Hetero. P. Type | Hetero.P. Amount | RecA Amount | |
|-----|-----------------|-------------------|--------------------|---------------------|----------------|-------|
| 5 | 1 | 30 bases | 150ng | - | - | 4.5μg |
| | 2 | 30 bases | 5ng | lambda DNA fragment | 145ng | 4.5μg |
| | 3 | 60 bases | 150ng | - | - | 4.5μg |
| | 4 | 60 bases | 5ng | lambda DNA fragment | 145ng | 4.5μg |
| | 5 | 275 bases | 150ng | - | - | 4.5μg |
| | 6 | 275 bases | 5ng | lambda DNA fragment | 145ng | 4.5μg |

10

Table 8

| | No. | Number of Amp ^r colonies | Specificity (%) |
|----|-----|--|-----------------|
| 15 | 1 | 1,778 | 6.6 |
| | 2 | 248 | 50.0 |
| | 3 | 1,119 | 3.3 |
| | 4 | 115 | 60.0 |
| | 5 | 7,871 | 0.0 |
| | 6 | 130 | 66.7 |

From the above results, in the screening of target cDNA from an actual cDNA library, it is confirmed that the specificity is greatly improved by the addition of heterologous probe.

The present invention has made it possible to target, enrich, detect and/or isolate the double-stranded target DNA molecule existing in the DNA sample in extremely small amount with high efficiency and specificity by using the short-strand homologous probe and heterologous probe. The present invention is expected to be applied to isolating or cloning a target gene from a cDNA or a genomic DNA mixture; screening for a target gene in various gene libraries, amplifying a target DNA sequence or mapping genes using RecA like recombinases; performing sequence-specific modification of or cleaving a target DNA using oligonucleotides; and in-situ

hybridizing and in vivo gene targeting performed in living cells using RecA-like recombinase.

SUMMARY OF SEQUENCES

- SEQ ID NO:1 is a double-stranded DNA fragment of p53 cDNA.
- 5 SEQ ID NO:2 is a double-stranded DNA fragment of p53 cDNA.
- SEQ ID NO:3 is a double-stranded DNA fragment of lambda phage genomic DNA sequence.
- SEQ ID NO:4 is a double-stranded DNA fragment of lambda phage genomic DNA sequence.
- 10 SEQ ID NO:5 is a double-stranded DNA fragment of lambda phage genomic DNA sequence.
- SEQ ID NO:6 is a single-stranded DNA probe corresponding to a region of p53 cDNA.
- SEQ ID NO:7 is a single-stranded DNA probe corresponding to a region of
- 15 lambda phage genomic DNA.
- SEQ ID NO:8 is a double-stranded DNA fragment of p53 cDNA.
- SEQ ID NO:9 is a double-stranded DNA fragment of p53 cDNA.
- SEQ ID NO:10 is a double-stranded DNA fragment of p53 cDNA.
- SEQ ID NO:11 is an oligonucleotide primer for p53 cDNA.
- 20 SEQ ID NO:12 is an oligonucleotide primer for p53 cDNA.
- SEQ ID NO:13 is an oligonucleotide primer for p53 cDNA.
- SEQ ID NO:14 is an oligonucleotide primer for p53 cDNA.
- SEQ ID NO:15 is an oligonucleotide primer for p53 cDNA.
- SEQ ID NO:16 is an oligonucleotide primer for p53 cDNA.
- 25 SEQ ID NO:17 is an oligonucleotide primer for p53 cDNA.
- SEQ ID NO:18 is an oligonucleotide primer for lambda phage genomic DNA.
- SEQ ID NO:19 is an oligonucleotide primer for lambda phage genomic DNA.
- SEQ ID NO:20 is an oligonucleotide primer for lambda phage genomic DNA.
- SEQ ID NO:21 is an oligonucleotide primer for lambda phage genomic DNA.
- 30 SEQ ID NO:22 is an oligonucleotide primer for lambda phage genomic DNA.

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The foregoing is meant to illustrate, but not to limit, the scope of the invention. Indeed, those of ordinary skill in the art can readily envision and produce further embodiments, based on the teachings herein, without undue experimentation.

5 The present invention may be embodied in other specific forms without departing from its essential characteristics. The described embodiment is to be considered in all respects as illustrative only and not as restrictive. The scope of the invention is, therefore, indicated by the appended claims rather than by the foregoing description. All changes which come within the meaning and range of the equivalence of the claims are to be embraced within the scope of the claims.

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SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: DAIKIN INDUSTRIES LTD.

(ii) TITLE OF INVENTION: METHODS FOR TARGETING, ENRICHING,
 5 DETECTING AND/OR ISOLATING TARGET NUCLEIC ACID SEQUENCE
 USING RECA-LIKE RECOMBINASE

(iii) FILE REFERENCE: D2-802PCT

(iv) CURRENT APPLICATION NUMBER:

(v) CURRENT FILING DATE:

(vi) EARLIER APPLICATION NUMBER: JP 8-229061, JP 8-347090

(vii) EARLIER FILING DATE: August 29, 1996, December 26, 1996

(viii) NUMBER OF SEQUENCES: 22

(2) INFORMATION FOR SEQ ID NO:1:

(i) SEQUENCE CHARACTERISTICS:

15 (A) LENGTH: 275 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

20 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CCTTGCCGTC CCAAGCAATG GATGATTGA TGCTGTCCC GGACGATATT GAACAATGGT 60
 TCACTGAAGA CCCAGGTCCA GATGAAGCTC CCAGAATGCC AGAGGCTGCT CCCCGCGTGG 120
 CCCCTGCACC AGCAGCTCCT ACACCGGCGG CCCCTGCACC AGCCCCCTCC TGGCCCCCTGT 180
 CATCTTCTGT CCCTTCCCAG AAAACCTACC AGGGCAGCTA CGGTTTCCGT CTGGGCTTCT 240
 25 TGCATTCTGG GACAGCCAAG TCTGTGACTT GCACG 275

(2) INFORMATION FOR SEQ ID NO:2:

(i) SEQUENCE CHARACTERISTICS:

30 (A) LENGTH: 60 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:

AGCTACGGTT TCCGTCTGGG CTTCTTGCAT TCTGGGACAG CCAAGTCTGT GACTTGCACG 60

35 (2) INFORMATION FOR SEQ ID NO:3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 500 base pairs
 (B) TYPE: nucleic acid

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(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

5 GATGAGTTCG TGTCCGTACA ACTGGCGTAA TCATGGCCCT TCGGGGCCAT TGTITCTCTG 60
 TGGAGGAGTC CATGACGAAA GATGAACTGA TTGCCCGTCT CCGCTCGCTG GGTGAACAAC 120
 TGAACCGTGA TGTCAGCCTG ACGGGGACGA AAGAAGAACT GGCGCTCCGT GTGGCAGAGC 180
 TGAAAGAGGA GCTTGATGAC ACGGATGAAA CTGCCGGTCA GGACACCCCT CTCAGCCGGG 240
 AAAATGTGCT GACCGGACAT GAAAATGAGG TGGGATCAGC GCAGCCGGAT ACCGTGATTC 300
 10 TGGATACGTC TGAAC TGGTC ACGGTCGTGG CACTGGTGAA GCTGCATACT GATGCACTTC 360
 ACGCCACGCG GGATGAACCT GTGGCATTTC TGCTGCCGGG AACGGCGTTT CGTGTCTCTG 420
 CCGGTGTGGC AGCCGAAATG ACAGAGCGCG GCCTGGCCAG AATGCAATAA CGGGAGGCGC 480
 TGTGGCTGAT TTCGATAACC 500

(2) INFORMATION FOR SEQ ID NO:4:

15 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1393 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

20 (ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:

GCGGCACGGA GTGGAGCAAG CGTGACAAGT CCACGTATGA CCGGACCGAC GATATCGAAG 60
 CCTACGCGCT GAACGCCAGC GGTGTGGTGA ATATCATCGT GTTCGATCCG AAGGGCTGGG 120
 CGCTGTTCCG TTCCTTCAAA GCCGTCAAGG AGAAGCTGGA TACCGTCTGT GGCTCTAATT 180
 25 CCGAGCTGGA GACAGCGGTG AAAGACCTGG GCAAGCGGT GTCTATAAG GGGATGTATG 240
 GCGATGTGGC CATCGTCGTG TATCCGGAC AGTACGTGGA AACGGCGTC AAAAAAGAACT 300
 TCCTGCCGGA CAACACGATG GTGCTGGGGA AACTCAGGC ACGCGGTCTG CGCACCTATG 360
 GCTGCATTCA GGATGCGGAC GCACAGCGCG AAGGCAATTA CGCTCTGCC CGTTACCCGA 420
 AAAACTGGGT GACCACCGGC GATCCGGCGC GTGAGTTCAC CATGATTGAG TCAGCACCGC 480
 30 TGATGCTGCT GGCTGACCCT GATGAGTTCG TGTCCGTACA ACTGGCGTAA TCATGGCCCT 540
 TCGGGGCCAT TGTITCTCTG TGGAGGAGTC CATGACGAAA GATGAACTGA TTGCCCGTCT 600
 CCGCTCGCTG GGTGAACAAC TGAACCGTGA TGTCAGCCTG ACGGGGACGA AAGAAGAACT 660
 GGCGCTCCGT GTGGCAGAGC TGAAAGAGGA GCTTGATGAC ACGGATGAAA CTGCCGGTCA 720
 GGACACCCCT CTCAGCCGGG AAAATGTGCT GACCGGACAT GAAAATGAGG TGGGATCAGC 780
 35 GCAGCCGGAT ACCGTGATTC TGGATACGTC TGAAC TGGTC ACGGTCGTGG CACTGGTGAA 840
 GCTGCATACT GATGCACTTC ACGCCACGCG GGATGAACCT GTGGCATTTC TGCTGCCGGG 900
 AACGGCGTTT CGTGTCTCTG CCGGTGTGGC AGCCGAAATG ACAGAGCGCG GCCTGGCCAG 960
 AATGCAATAA CGGGAGGCGC TGTGGCTGAT TTCGATAACC TGTTCGATGC TGCCATTGCC 1020
 CGGCGCGATG AAACGATACG CGGGTACATG GGAACGTGAG CCACCAATAC ATCCGGTGAG 1080
 40 CAGTCAGGTG CCGTGATACG TGGTGTTCCT GATGACCCTG AAAATATCAG CTATGCCGGA 1140
 CAGGGCGTGC GCGTTGAAGG CTCCAGCCCG TCCCTGTTTG TCCGGACTGA TGAGGTGCGG 1200

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CAGCTGCGGC GTGGAGACAC GCTGACCATC GGTGAGGAAA ATTTCTGGGT AGATCGGGTT 1260
 TCGCCGGATG ATGGCGGAAG TTGTCATCTC TGGCTTGGAC GGGGCGTACC GCCTGCCGTT 1320
 AACCGTCGCC GCTGAAAGGG GGATGTATGG CCATAAAAGG TCTTGAGCAG GCCGTTGAAA 1380
 ACCTCAGCCG TAT 1393

5 (2) INFORMATION FOR SEQ ID NO:5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 308 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double

10 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

AGGTGCGGTG ATACGTGGTG TTTTGTATGA CCTTGAAAAT ATCAGCTATG CCGGACAGGG 60
 CGTGCGCGTT GAAGGCTCCA GCGCGTCCCT GTTTGTCCGG ACTGATGAGG TGCGGCAGCT 120
 15 GCGGCGTGGA GACACGCTGA CCATCGGTGA GGAAAATTTC TGGGTAGATC GGGTTTCGCC 180
 GGATGATGGC GGAAGTTGTC ATCTCTGGCT TGGACGGGGC GTACCGCTG CCGTTAACCG 240
 TCGCCGCTGA AAGGGGGATG TATGGCCATA AAAGGTCTTG AGCAGGCCGT TGA AACCTC 300
 AGCCGTAT 308

20 (2) INFORMATION FOR SEQ ID NO:6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 30 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

25 (ii) MOLECULE TYPE: cDNA to mRNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

CCTTGCCGTC CCAAGCAATG GATGATTTGA 30

(2) INFORMATION FOR SEQ ID NO:7:

(i) SEQUENCE CHARACTERISTICS:

- 30 (A) LENGTH: 60 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: DNA (genomic)

35 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

AACGGCGTTT CGTGTCTCTG CCGGTGTGGC AGCCGAAATG ACAGAGCGCG GCCTGGCCAG 60

(2) INFORMATION FOR SEQ ID NO:8:

(i) SEQUENCE CHARACTERISTICS:

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(A) LENGTH: 1317 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

5 (ii) MOLECULE TYPE: cDNA to mRNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:

GTCTAGAGCC ACCGTCCAGG GAGCAGGTAG CTGCTGGGCT CCGGGGACAC TTTGCGTTCCG 60
 GGCTGGGAGC GTGCTTTCCA CGACGGTGAC ACGCTTCCCT GGATTGGCAG CCAGACTGCC 120
 TTCCGGGTCA CTGCCATGGA GGAGCCGCAG TCAGATCCTA GCGTCGAGCC CCCTCTGAGT 180
 10 CAGGAAACAT TTTCAGACCT ATGGAACTA CTTCTGAAA ACAACGTTCT GTCCCCCTTG 240
 CCGTCCCAAG CAATGGATGA TTTGATGCTG TCCCCGGACG ATATTGAACA ATGGTTCACT 300
 GAAGACCCAG GTCCAGATGA AGCTCCAGA ATGCCAGAGG CTGCTCCCG CGTGGCCCCCT 360
 GCACCAGCAG CTCCTACACC GCGGGCCCCCT GCACCAGCCC CCTCTGGCC CCTGTCATCT 420
 TCTGTCCCTT CCCAGAAAAC CTACCAGGGC AGCTACGGT TCCGTCTGGG CTCTTTGCAT 480
 15 TCTGGGACAG CCAAGTCTGT GACTTGCACG TACTCCCTG CCTTCAACAA GATGTTTTGC 540
 CAACTGGCCA AGACCTGCCC TGTGCAGCTG TGGGTGAT CCACACCCC GCCCGGCACC 600
 CCGCTCCGCG CCATGGCCAT CTACAAGCAG TCACAGCACA TGACGGAGGT TGTGAGGCGC 660
 TGCCCCCACC ATGAGCGCTG CTCAGATAGC GATGGTCTGG CCCCTCCTCA GCATCTTATC 720
 CGAGTGGAAG GAAATTTGCG TGTGGAGTAT TTGGATGACA GAAACACTTT TCGACATAGT 780
 20 GTGGTGGTGC CCTATGAGCC GCCTGAGGTI GGCTCTGACT GTACCACCAT CCACTACAAC 840
 TACATGTGTA ACAGTTCCTG CATGGGCGGC ATGAACCAGA GGCCCATCCT CACCATCATC 900
 ACACTGGAAG ACTCCAGTGG TAATCTACTG GGACGGAACA GCTTTGAGGT GCGTGTITGT 960
 GCCTGTCTG GGAGAGACCG GCGCACAGAG GAAGAGAATC TCCGCAAGAA AGGGGAGCCT 1020
 CACCACGAGC TGCCCCCAGG GAGCACTAAG CGAGCACTGC CCAACAAAC CAGCTCCTCT 1080
 25 CCCCAGCCAA AGAAGAAACC ACTGGATGGA GAATATTTCA CCTTCAGAT CCGTGGGCGT 1140
 GAGCGCTTCG AGATGTTCCG AGAGCTGAAT GAGGCCITGG AACTCAAGGA TGCCAGGCT 1200
 GGGAAAGGAGC CAGGGGGGAG CAGGGCTCAC TCCAGCCACC TGAAGTCCAA AAAGGGTCAG 1260
 TCTACCTCCC GCCATAAAAA ACTCATGTTT AAGACAGAAG GGCTGACTC AGACTGA 1317

(2) INFORMATION FOR SEQ ID NO:9:

30 (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 410 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: double

(D) TOPOLOGY: linear

35 (ii) MOLECULE TYPE: cDNA to mRNA

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:

CCTTGCCGTC CCAAGCAATG GATGATTTGA TGCTGTCCCC GGACGATATT GAACAATGGT 60
 TCACTGAAGA CCCAGGTCCA GATGAAGCTC CCAGAATGCC AGAGGCTGCT CCCCAGGTGG 120
 CCCCTGCACC AGCAGCTCCT ACACCGGCGG CCCCTGCACC AGCCCCCTCC TGCCCCCTGT 180
 40 CATCTTCTGT CCTTCCCAG AAAACCTACC AGGGCAGCTA CGGTTTCCGT CTGGGCTTCT 240
 TGCATTCTGG GACAGCCAAG TCTGTGACTT GCACGTACTC CCCTGCCCTC AACAAGATGT 300

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TTTGCCAACT GGCCAAGACC TGCCCTGTGC AGCTGTGGGT TGATTCCACA CCCCCGCCCCG 360
GCACCCGCGT CCGCGCCATG GCCATCTACA AGCAGTCACA GCACATGACG 410

(2) INFORMATION FOR SEQ ID NO:10:

(i) SEQUENCE CHARACTERISTICS:

- 5 (A) LENGTH: 779 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: double
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA to mRNA

10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

CCTTGCCGTC CCAAGCAATG GATGATTTGA TGCTGTCCCC GGACGATATT GAACAATGGT 60
TCACTGAAGA CCCAGGTCCA GATGAAGCTC CCAGAATGCC AGAGGCTGCT CCCC GCGTGG 120
CCCCTGCACC AGCAGCTCCT ACACCGGCGG CCCCTGCACC AGCCCCCTCC TGGCCCCCTGT 180
CATCTTCTGT CCCTTCCCAG AAAACCTACC AGGGCAGCTA CGGTTTCCGT CTGGGCTTCT 240
15 TGCATTCTGG GACAGCCAAG TCTGTGACTT GCACGTACTC CCCTGCCCTC AACAAAGATGT 300
TTTGCCAACT GGCCAAGACC TGCCCTGTGC AGCTGTGGGT TGATTCCACA CCCCCGCCCCG 360
GCACCCGCGT CCGCGCCATG GCCATCTACA AGCAGTCACA GCACATGACG GAGGTTGTGA 420
GGCGCTGCCC CCACCATGAG CGCTGCTCAG ATAGCGATGG TCTGGCCCTT CCTCAGCATC 480
TTATCCGAGT GGAAGGAAAT TTGCGTGTGG AGTATTGGA TGACAGAAAC ACTTTTCGAC 540
20 ATAGTGTGGT GGTGCCCTAT GAGCCGCTG AGGTTGGCTC TGACTGTACC ACCATCCACT 600
ACAACTACAT GTGTAACAGT TCCTGCATGG GCGGCATGAA CCAGAGGCCC ATCCTCACCA 660
TCATCACACT GGAAGACTCC AGTGGTAATC TACTGGGACG GAACAGCTTT GAGGTGCGTG 720
TTTGTGCCTG TCCTGGGAGA GACCGGCGCA CAGAGGAAGA GAATCTCCGC AAGAAAGGG 779

(2) INFORMATION FOR SEQ ID NO:11:

25 (i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

30 (ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:11:

CCTTGCCGTC CCAAGCAATG GATGA 25

(2) INFORMATION FOR SEQ ID NO:12:

(i) SEQUENCE CHARACTERISTICS:

- 35 (A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

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- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:
CGTGCAAGTC ACAGACTTGG CTGTC 25
- (2) INFORMATION FOR SEQ ID NO:13:
- (i) SEQUENCE CHARACTERISTICS:
- 5 (A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
- 10 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:
AGCTACGGTT TCCGTCTGGG CTTCT 25
- (2) INFORMATION FOR SEQ ID NO:14:
- (i) SEQUENCE CHARACTERISTICS:
- 15 (A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:
- 20 CGTCATGTGC TGTGACTGCT TGTAG 25
- (2) INFORMATION FOR SEQ ID NO:15:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
25 (C) STRANDEDNESS: single
(D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:
- CCCTTTCTTG CGGAGATTCT CTTCC 25
- 30 (2) INFORMATION FOR SEQ ID NO:16:
- (i) SEQUENCE CHARACTERISTICS:
- (A) LENGTH: 25 base pairs
(B) TYPE: nucleic acid
(C) STRANDEDNESS: single
35 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: other nucleic acid
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:

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GTCTAGAGCC ACCGTCCAGG GAGCA

25

(2) INFORMATION FOR SEQ ID NO:17:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

5

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:

10

TCAGTCTGAG TCAGGCCCTT CTGTC

25

(2) INFORMATION FOR SEQ ID NO:18:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

15

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:

GATGAGTTCG TGTCCGTACA ACTGG

25

20

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

25

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:

GGTTATCGAA ATCAGCCACA GCGCC

25

30

(2) INFORMATION FOR SEQ ID NO:20:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

35

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:

GCGGCACGGA GTGGAGCAAG CGTGA

25

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(2) INFORMATION FOR SEQ ID NO:21:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

5 (C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:

ATACGGCTGA GGTTTTCAAC GGCCT

25

10 (2) INFORMATION FOR SEQ ID NO:22:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 25 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

15 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: other nucleic acid

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:

AGGTGCGGTG ATACGTGGTG TTTT

25

CLAIMS

1. A method for targeting, enriching, detecting and/or isolating a double-stranded nucleic acid target sequence in a sample by using a RecA-like recombinase, comprising the steps of:
 - 5 (a) providing at least one type of recombinase, at least one type of homologous probe, and at least one type of heterologous probe, and
 - (b) mixing the recombinase, the homologous probe, and the heterologous probe with the double-stranded nucleic acid target sequence in the sample,wherein the specificity of the homologous pairing and/or strand exchange between
10 the double-stranded target nucleic acid sequence and the homologous nucleic acid probe is increased due to the addition of the heterologous nucleic acid probe.
2. The method of claim 1, wherein the step (b) further comprises the steps of:
 - (a) reacting the homologous probe and the heterologous probe with the recombinase respectively for coating the probes with the recombinase, and
 - 15 (b) applying the recombinase-coated homologous probe and the heterologous probe to the double-stranded nucleic acid target sequence in the sample.
3. The method of claim 1, wherein the double-stranded nucleic acid target sequence is double-stranded target DNA.
4. The method of claim 1, wherein the step (a) provides at least two types of
20 homologous probes.
5. The method of claim 4, wherein the homologous probes are two types of homologous probes which are sufficiently complementary to each other.
6. The method of claim 1, wherein the homologous probe is at least 15 nucleic acid residues in length.
- 25 7. The method of claim 6, wherein the homologous probe is about 15 to 2000 nucleic acid residues in length.

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8. The method of claim 1, wherein the homologous probe comprises a label or a ligand.
9. The method of claim 1, wherein the homologous probe is a double-stranded DNA probe.
- 5 10. The method of claim 1, wherein the homologous probe is a single-stranded DNA probe.
11. The method of claim 1, wherein the heterologous probe is about 15 bases or longer.
- 10 12. The method of claim 1, wherein the heterologous probe is a single-stranded DNA probe.
13. The method of claim 1, wherein the heterologous probe is a RNA probe.
14. The method of claim 1, wherein the weight ratio of the homologous probe to the heterologous probe is about 1:1 to about 1:500.
- 15 15. The method of claim 1, wherein the RecA-like recombinase is derived from a prokaryote.
16. The method of claim 15, wherein the RecA-like recombinase is derived from *Escherichia coli*.
17. The method of claim 16, wherein the RecA-like recombinase is RecA.
- 20 18. A method for targeting, enriching, detecting and/or isolating a double-stranded nucleic acid target sequence in a sample by using a RecA-like recombinase, comprising the steps of:

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(a) providing a homologous probe which has a label or a ligand and which is bound to at least one type of recombinase, and a heterologous probe bound to at least one type of recombinase;

5 (b) reacting the recombinase-coated homologous probe and the recombinase-coated heterologous probe with the double-stranded nucleic acid target sequence under conditions which promote the formation of a complex of the double stranded nucleic acid target sequence and the homologous probe in the reaction mixture;

10 (c) contacting the reaction mixture with a solid support designed to selectively bind the complex of the double-stranded nucleic acid target sequence and the homologous probe;

(d) removing non-bound molecules from the solid support; and

(e) treating the solid support to release the double-stranded nucleic acid target sequence which is selectively bound to the solid support through the complex.

15 19. The method of claim 18, wherein the double-stranded nucleic acid target sequence is double-stranded target DNA.

20. The method of claim 19, wherein the sample is a cDNA or a genomic DNA library, and the double-stranded target DNA is inserted to a vector capable of transforming or transfecting a suitable bacterial host.

21. The method of claim 20 further comprising the steps of:

20 (a) providing suitable host cells which are capable of being transformed or transfected by the double-stranded target DNA inserted into a vector;

(b) transforming or transfecting the host cells with the target DNA fraction released from the solid phase; and

(c) selecting transformed or transfected cells containing said target DNA.

25 22. The method of claim 19, wherein the sample is a clinical specimen containing a mixture of cDNA or genomic DNA

23. The method of claim 18, wherein the label or ligand is biotin or digoxigenin.

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24. The method of claim 18, wherein the solid phase comprises magnetic or paramagnetic beads surfacely bound to avidin, streptavidin, or anti-digoxigenin antibody.

25. The method of claim 18, wherein at least two types of homologous probes are provided.

5 26. The method of claim 25, wherein two types of homologous probes which are sufficiently complementary to each other are provided.

27. The method of claim 18, wherein the RecA-like recombinase is derived from prokaryote.

10 28. The method of claim 27, wherein the RecA-like recombinase is derived from *Escherichia coli*.

29. The method of claim 18, wherein the heterologous probe is at least 15 nucleic acid residues in length.

15 30. The method of claim 18, wherein the homologous probe is at least 15 nucleic acid residues in length.

31. The method of claim 18, wherein the weight ratio of the homologous probe to the heterologous probe is about 1:1 to about 1:500.

20 32. A method for detecting a double-stranded nucleic acid target sequence in a sample of cells by in situ hybridization method using a RecA-like recombinase, comprising the steps of

(a) providing a homologous probe and a heterologous probe, wherein the probes are bound to the recombinase;

25 (b) adding the recombinase-coated homologous probe and heterologous probe to the sample of cells under a condition to promote the reaction of the homologous probe with the double-stranded nucleic acid target sequence; and

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(c) removing the probes which have not reacted with the double-stranded nucleic acid target sequence.

33. The method of claim 32, wherein the weight ratio of the homologous probe to the heterologous probe is about 1:1 to about 1:500.

5 34. The method of claim 32, wherein at least two types of homologous probes are provided.

35. The method of claim 34, wherein two types of homologous probes which are sufficiently complementary to each other are employed.

10 36. The method of claim 32, wherein the heterologous probe is of 15 bases or longer.

37. The methods of claims 2, 18 or 32, wherein the homologous and heterologous probes are bound to recombinase in the presence of a co-factor.

38. The method of claim 37, wherein the RecA-like recombinase is derived from *Escherichia coli*.

15 39. The method of claim 37, wherein the cofactor is selected from a group consisting of GTP γ S, a mixture of ATP γ S and ADP, a mixture of ADP and AlF $_4^-$ (aluminum nitrate and sodium fluoride), a mixture of ATP and AlF $_4^-$ (aluminum nitrate and sodium fluoride), a mixture of dADP and AlF $_4^-$ (aluminum nitrate and sodium fluoride), or a mixture of dATP and AlF $_4^-$ (aluminum nitrate and sodium fluoride).

20

40. The method of claim 39, wherein the cofactor is GTP γ S,

41. A method for targeting a double-stranded nucleic acid target sequence in a sample of living cells by an in vivo gene targeting method using a RecA-like recombinase, comprising the steps of:

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(a) providing a recombinase coated homologous probe and a recombinase coated heterologous probe;

(b) introducing the homologous probe and the heterologous probe into the living cells;

5 (c) incubating the living cells containing the homologous probe and the heterologous probe for a sufficient period of time to allow the homologous probe to be transformed into the genome of the cells.

42. The method of claim 41, wherein the weight ratio of the homologous probe to the heterologous probe is about 1:1 to about 1:500.

10 43. A kit for enriching, detecting and/or isolating of a double-stranded nucleic acid target sequence, comprising a RccA-like recombinase, a cofactor, a heterologous probe, a solid phase designed to trap a complex of a double-stranded nucleic acid target sequence and a homologous probe having a label or ligand, and washing buffer.

15 44. The method of claim 43, wherein the weight ratio of the homologous probe to the heterologous probe is about 1:1 to about 1:500.

INTERNATIONAL SEARCH REPORT

Int'l Application No

PCT/JP 97/03019

A. CLASSIFICATION OF SUBJECT MATTER
IPC 6 C12Q1/68

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C12Q

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

| Category | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|----------|---|-----------------------|
| X | EP 0 687 738 A (DAIKIN IND LTD) 20 December 1995 page 8, left column, claim 37 see the whole document --- | 1-44 |
| A | US 5 468 620 A (CALHOUN CORNELIA) 21 November 1995 cited in the application see the whole document --- | 1-44 |
| A | WO 93 05177 A (DAIKIN IND LTD) 18 March 1993 cited in the application see the whole document --- | 1-44 |
| | --- -/-- | |

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex

* Special categories of cited documents :

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Date of the actual completion of the international search

11 December 1997

Date of mailing of the international search report

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INTERNATIONAL SEARCH REPORT

International Application No

PCT/JP 97/03019

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

| Category | Citation of document, with indication, where appropriate, of the relevant passages | Relevant to claim No. |
|----------|---|-----------------------|
| A | WO 93 05178 A (DAIKIN IND LTD) 18 March 1993 cited in the application see the whole document ---- | 1-44 |
| A | TEINTZE ET AL.: "RecA assisted rapid enrichment of specific clones from model DNA libraries" BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, vol. 211, no. 3, 1995, pages 804-811, XP002049831 cited in the application see the whole document ----- | 1-44 |

INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/JP 97/03019

| Patent document cited in search report | Publication date | Patent family member(s) | Publication date |
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